

RESEARCH **Open Access** 

# Effect of difference doses of Newcastle disease vaccine immunization on growth performance, plasma variables and immune response of broilers

Xiaofei Wang<sup>1,2</sup>, Qinqin Zhou<sup>1</sup>, Jing Shen<sup>1</sup>, Junhu Yao<sup>1\*</sup> and Xiaojun Yanq<sup>1\*</sup>

#### **Abstract**

Background: As a member of the Paramyxoviridae group, Newcastle disease virus (NDV) is the key causative agent of Newcastle disease (ND) that attacks chickens, turkeys and other avian birds. Surviving birds showed lower feed utilization, growth performance or egg production, which results in severe economic losses. The purpose of this study was to determine the effect of different doses of NDV immunization on growth performance, plasma variables and immune response of broiler chickens.

Methods: A total of 480 one-day-old Arbor Acres broilers were randomly administrated with 0, 4, 6 or 8 doses of NDV at 12 d and 28 d, respectively. Each group consisted of ten replicates with 12 birds each. Growth performance and organ weight were recorded. Plasma concentration of glucose, total protein, cholesterol, triglycerides and nonesterified fatty acid was determined using commercial kits. The concentration of plasma corticosterone and insulin was measured using commercially available radio immune assay kits. Serum antibody titer and peripheral blood lymphocyte proliferation were also recorded.

Results: The results showed that NDV decreased body weight gain (BWG), and increased Feed: Gain ratio at 1-21 d at all doses (P < 0.05). Plasma insulin concentration was lower in all immunization groups after the first immunization at 12 d (P < 0.01). The rest of the plasma indexes were not affected by NDV immunization, including glucose, total protein, cholesterol, triglycerides, nonesterified fatty acid, heterophil/lymphocyte ratio, as well as the proliferation of peripheral blood lymphocyte (P > 0.05). Compared with the control group, NDV treatment elevated NDV antibody titer at 10 d after the first inoculation (P < 0.05), and at d 5, 9 and 13 after the second inoculation (P < 0.05). Repeated NDV inoculation had no deleterious impacts on body composition at 42 d, and nutrient accretion rates at 8-42 d (P > 0.05).

**Conclusions:** In conclusion, NDV challenge decreased BWG and feed efficiency in earlier stage of growth. However, NDV treatment at 6 doses down-regulated the Feed: Gain ratio by 6.36 % throughout the whole growing period. These data suggest that appropriate lower doses of NDV inoculation increase feed efficiency of broiler chickens.

**Keywords:** Broiler, Immune response, Metabolism, Newcastle disease vaccine

<sup>\*</sup> Correspondence: yaojunhu2004@sohu.com; yangxj@nwsuaf.edu.cn <sup>1</sup>College of Animal Science and Technology, Northwest A&F University, Yangling, Shaanxi, China Full list of author information is available at the end of the article



### **Background**

As a member of the Paramyxoviridae group, Newcastle disease virus (NDV) is the key causative agent of Newcastle disease (ND) that attacks chickens, turkeys and other avian birds [1]. Surviving birds showed impaired growth, altered feed utilization, reduced egg production and resulting in severe economic losses [2]. To alleviate its detrimental effect on growth rate and efficiency of gain, NDV inoculation has been used to protect the chick from ND, and this is one of the challenges that broiler faced throughout their life.

Immune stress as well as other forms of stresses is directly or indirectly translated to the body by the following two common ways: hypothalamic-pituitary-adrenal axis, resulting in the release of corticotropin-releasing hormone, adrenocorticotropic hormone and glucocorticoid and the sympathetic-adrenal medullary axis, resulting in the release of catecholamines [3]. It is reported that continuous delivery of adrenocorticotropic hormone increased plasma corticosterone, glucose, cholesterol and total protein concentration [4]. Elevated corticosterone concentration resulted in many deleterious effects on broiler performance. Corticosterone treatment affected the preference and appetite of broilers [5, 6], and had inversely proportional to body weight [7]. Corticosterone induced gluconeogenesis, protein catabolism and fat deposition [8, 9]. Insulin signalling in chicken liver and muscle was also impaired by corticosterone through different mechanisms [10].

Our previous study showed that routine vaccine immunization decreased feed intake, body weight gain (BWG), nutrients digestibility and accompanied by markedly enhanced plasma IgG concentration [11]. However, sheep red blood cells immunization dose not induce any difference on feed intake, body weight, rate of gain and the metabolizability of gross energy in pullets [12]. Growth performance was not affected, and immune response required very little nutrient cost by a single or limited amount of immunization in domesticated fowl [13]. Moreover, in modern poultry industry, vaccines were administered in large doses at a high frequency to reduce the mortality, especially when using oral vaccines [14]. The effect of high level vaccination on immune status, energy and nutrients metabolism in broiler chickens was not fully elucidated. So in the present study, we used vaccination itself as a stressor to investigate the effect of different levels of NDV on the performance, metabolism, immune responses and carcass characteristics of Arbor Acres broilers for better understanding of immune stress on broiler chickens.

### **Methods**

#### Husbandry and treatments

All protocols were approved by the animal care committee of Northwest A&F University. One-day-old Arbor

Acres broiler chicks were obtained from Shaanxi Zhengda Commercial Hatchery (Xi'an, China) and housed in a temperature controlled room. Ambient temperature was controlled at 35–33 °C for the first week and declined 2 °C each week until it reached 24 °C. Feed and water were supplied *ad libitum*, and continuous light was provided for the duration of the experiments. All diets were formulated to meet or exceed the requirements of National Research Council [15] (Additional file 1: Table S1). On d 8,480 broilers were randomly allotted to four groups, with six replicates and 20 birds each. The initial body weight (BW) was not significantly different.

Newcastle disease virus thermo-stable live vaccine (La Sota Srain, yellowish spongy solid, Nanjing Tech-bank Bio-industry Co., Ltd, China) was dissolved using sterilized saline. The chickens in good health were administrated with 4, 6 and 8 doses of the vaccine at 12 d and 28 d by eye-drop and drinking water, respectively (abbreviated as 4 doses, 6 doses and 8 doses groups, respectively).

#### **Growth performance**

The BW and feed intake were recorded each week. The average BW and feed intake were calculated at 12 d and 42 d, respectively.

### Blood collection and organ weight

On d 12 and 28, one bird from each replicate was sampled after BW measurement. Approximately 8 mL blood sample was collected and transferred to a heparinized centrifuge tube. One part of the blood was used for heterophil/lymphocyte ratio and lymphocyte proliferation assay, and the other part was centrifuged at 2,000 g for 10 min to obtain plasma for later biochemical analysis. On d 28, the birds were killed by cervical dislocation after blood sampling for the weight measurement of bursa of fabricius, spleen and thymus. Relative weights of each organ were calculated.

#### **Blood parameter measurement**

On d 12 and 28, the heterophil/lymphocyte ratio was determined according to Zhang et al. [16]. In brief, 5  $\mu$ L blood was used to make a smear on a clean glass slide. The smears were stained with Wright-Giemsa after dried. The cell numbers were counted under a microscope at a magnification of ×100 with oil immersion by a trained person. In each visual section, a cumulative total of 100 cells including heterophils and lymphocytes were counted. The heterophils/lymphocytes ratios were then calculated. The concentration of plasma corticosterone and insulin was measured using available radio immune assay kits for corticosterone (ICN Pharmaceuticals, Costa Mesa) and insulin (Beijing North Institute of Biological Technology, Beijing, China), respectively. The

concentration of plasma glucose, total protein, cholesterol, triglycerides and nonesterified fatty acid was determined with commercial kits according to the manufactures' instructions (Nanjing Jiancheng Bioengineering Institute, Nanjing, China).

#### Peripheral blood lymphocyte proliferation

The blood was collected at 0, 6, 12, 24 h, on d 2 and 4 after the second vaccination at 28 d to obtain the dynamic change regularity of lymphocyte proliferation. The T and B lymphocyte proliferation assay were determined according to Zhang and Guo [17] with some modifications. Lymphocyte separation medium (TBD science, Tianjin, China) was used to separate the lymphocyte. Lipopolysaccharide (LPS, 10  $\mu$ g/mL, Sigma) and Concanavalin A (45  $\mu$ g/mL, Sigma) were used to induce the proliferation of B lymphocyte and T lymphocyte, respectively [17]. The absorbance at 570 nm was then recorded using an automated ELISA microplate reader (Power Wave XS2, Bio Tek, USA). Stimulating index (SI) was calculated as follows: SI = OD<sub>570</sub> (experimental group)/OD<sub>570</sub> (blank control).

#### Serum antibody titers to NDV

Blood samples were collected from the wing vein at d 6, 10 and 14 after the first inoculation, and at d 5, 9 and 13 after the second inoculation of NDV. Serum samples were prepared and frozen at  $-80\,^{\circ}\text{C}$  until analyze.

NDV antibody quantification was used the hemagglutination inhibition method. The geometric mean titer was expressed as reciprocal log<sub>2</sub> values of the end point dilution.

#### Whole body sampling and determination

In order to determine the dry matter, nitrogen and ether-extractable lipid of the whole body, one bird from each replicate was killed and defeathered prior to the first inoculation (at 12 d) and at the end of the trial (at 42 d) to determine the initial and final body composition, respectively. The whole body was chopped into small pieces, and passed through a grinder (Electrical Mfg. Co., Changsha, China). All samples were lyophilized and reground for homogeneity. Dry matter was determined by drying at 105 °C for 24 h. Nitrogen and ether-extractable lipid were measured according to Association of Official Analytical Chemists [18] procedures.

#### Statistical analysis

The data were subjected to completely randomized design using the GLM procedure with SPSS 17.0. Significant differences amongst treatments were determined using Duncan's multiple range tests. P < 0.05 was considered statistically significant.

#### Results

#### **Growth performance**

Compared to control group, NDV treatment significantly lowered BWG for all doses at 1–21 d (P < 0.05, Additional file 1: Table S2), but the difference was not significant at 21–42 d and 1–42 d (P > 0.05). The Feed:Gain ratio was higher in NDV groups than control group at 1–21 d (P < 0.05). However, the ratio was significantly lower in 4 doses and 6 doses groups at 21–42 d (P < 0.05). Throughout the whole period, 6 doses treated group down-regulated the ratio by 6.36 % (P < 0.05). No significant difference was found amongst all treatments in feed intake for all experimental periods (P > 0.05). Relative weight of bursa of fabricius, thymus and spleen was not significantly different amongst treatments (data not shown).

#### Plasma parameters and heterophil/lymphocyte ratio

The effect of NDV inoculation on plasma biochemistry parameters were listed in Additional file 1: Table S3. After the first inoculation at 12 d, plasma corticosterone concentration was elevated in 8 doses group than that in 4 doses group (P < 0.05), and plasma insulin concentration was lower in all NDV treated groups (P < 0.05). However, no significant differences was found in concentration of plasma corticosterone and insulin amongst all groups after the second inoculation at 28 d (P > 0.05). The concentration of plasma glucose, total protein, cholesterol, triglycerides and nonesterified fatty acid was unaffected by the inoculation (P > 0.05). The heterophil/lymphocyte ratio tended to increase with the NDV inoculation dose, but the difference was not significant (P > 0.05).

### Antibody titer and lymphocyte proliferation

As shown in Additional file 1: Table S4, NDV immunization significantly elevated NDV antibody titer at 10 d after the first inoculation (P < 0.05), and at d 5, 9 and 13 after the second inoculation (P < 0.05). NDV antibody titer tended to increase at 14 d after the first inoculation, but did not reach significance (P > 0.05). The proliferation of T and B lymphocyte was not affected by the second inoculation at 28 d (P > 0.05). The stimulating index at 4 d returned to a concentration not different from 0 h after the inoculation (data not shown).

#### **Body nutrient content**

As shown in Additional file 1: Table S5, the body content of water, protein and lipid was not affected by NDV inoculation (P > 0.05). The nutrients accretion ratio was also unaffected by NDV immunization at 12–42 d (P > 0.05).

#### Discussion

Stress is consisted of different forms, and results in hormone releasing. Physiological stress is reported to reduce the BWG and feed conversion ratio [19, 20]. In the present study, NDV treatment reduced BWG and feed efficiency compared with the control group at 1-21 d. The absence of elevation in corticosterone after the second inoculation at 28 d might due to the adaptation of repeated NDV immunization. The BWG was higher and feed conversion was lower in both 4 doses and 6 doses groups than that in the control group, suggesting that a compensatory growth could exist at 21-42 d. Leeson and Zubair, and Lippens et al. also found that the birds suffered from feed restriction expressed equal or even higher energy intake and BWG in the whole phase [21, 22]. This can be explained by the metabolic adaptations and digestibility improvement [23, 24]. Moreover, 8 doses treatment did not decrease feed efficiency, indicating that vaccine should be manipulated at appropriate level.

It is well established that insulin is the key peptide hormone regulating the metabolism of carbohydrate and lipid in broilers. Insulin induces glucose disposal in adipose tissue and muscle, and also prevents the production of glucose in liver. In the present study, no significant change of plasma insulin and corticosterone was found following the second injection. These alterations in plasma hormones can well explain the aforementioned results of growth performance. Insulin resistance is one of the indicators of chronic stress [10]. However, lower plasma insulin concentration in NDV-treated birds was observed after the first inoculation, indicating that NDV treatment induced acute immune stress in our present study. Meanwhile, the plasma corticosterone concentration in NDV injected birds returned to the normal level after the second injection. Effect of stress on metabolism has been extensively investigated, and it is acceptable that stress causes a shift from anabolism to catabolism, and increases lipolysis and protein catabolism. In the present study, no significant change in concentration of plasma glucose, total protein, cholesterol, triglycerides and nonesterified fatty acid was observed. This might due to the sampling time or the NDV inoculation dose. More dynamic sampling time is needed for the further study.

As an important immune parameter, circulating heterophil/lymphocyte ratio is one of the most recognizable symptoms of stress in poultry [25]. Chicks treated with corticosterone showed depressed number of circulating lymphocytes [26]. In the present study, heterophil/lymphocyte ratio was slightly elevated in NDV-treated broilers following the second immunization at 28 d. Although the elevation was not significant, it also suggested that repeated NDV challenge can result in a certain immunosuppression.

The antibody titer and lymphocyte proliferation is corresponded to humoral and cellular immunity, respectively. Immune consequence is affected by immune type (immunoprotective, immunoregulatory/inhibitory, or immunopathological), duration and the aspects of immune system measured [27, 28]. In our study, the antibody titers against NDV were significantly enhanced in NDV-treated birds at 10 d after the first immunization, and at d 5, 9 and 13 after the second immunization, indicating that the humoral immune system was enhanced through repeated NDV challenge. Pathogen re-exposure often resulted in a more rapid and effective response than that of the first exposure due to immunologic memory [29]. No significant difference in lymphocyte proliferation in response to Concanavalin A or LPS was found in broilers between the control and NDV-administrated broilers at 28 d. We can speculate that NDV challenge enhanced the function or numbers of T helper cell, and subsequently modulated cytokine production and B cell activation [30].

Stressed broilers typically display a reduction in muscle accretion, and an increase in abdominal fat deposition [8]. However, in the present study, the whole body carcass composition and nutrient accretion rates were not different amongst all treatments, indicating that NDV immunization cannot alter broiler body composition.

#### **Conclusions**

NDV challenge reduced growth performance of broilers in earlier period after the first immunization, while appropriate lower NDV inoculation levels (4 or 6 doses) had beneficial effect on feed efficiency in later and the whole growing period. NDV antibody titer was increased at 10 d after the first NDV inoculation, and at d 5, 9 and 13 after the second NDV inoculation. These findings suggest that appropriate lower doses of NDV inoculation increase feed efficiency of broiler chickens.

#### **Additional file**

**Additional file 1: Table S1 to S5.** The ingredient composition and effect of Newcastle Disease vaccine (NDV) inoculation on growth performance, plasma parameters, Heterophil/Lymphocyte ratio, NDV antibody titer, body nutrient content and accretion ratio in broilers.

#### Competing interests

The authors declare that they have no competing interests.

#### Authors' contributions

XFW participated in the paper writing and whole body sampling and determination. QQZ carried out the whole *in vivo* trial, and participated in the paper writing, measurement of growth performance and organ weight. JS measured the blood parameter, peripheral blood lymphocyte proliferation and serum antibody titers. XJY and JHY made crucial contributions to the design of the whole trial and manuscript, and helped to modify the manuscript. All authors read and approved the final manuscript.

#### Acknowledgements

This research was supported by the Natural Science Foundation of China (No. 31001017, 31272464), the Program for New Century Excellent Talents (NCET-12-0476), the Science & Technological Project of Shaanxi Province, China (2011KTCQ02-02, 2011KTDZ02-01-01) and the Program for Shaanxi Youth Scientific Talents (2012KJXX-18). The authors are grateful to the staffs at Animal Nutrition and Feed Science of Northwest A&F University for their assistance in conducting the experiment.

#### **Author details**

<sup>1</sup>College of Animal Science and Technology, Northwest A&F University, Yangling, Shaanxi, China. <sup>2</sup>College of Science, Nanjing Agricultural University, Nanjing, Jiangsu 210095, China.

## Received: 21 October 2014 Accepted: 29 April 2015 Published online: 09 May 2015

#### References

- Mayo M. A summary of taxonomic changes recently approved by ICTV. Arch Virol. 2002;147:1655–6.
- Gallili GE, Ben-Nathan D. Newcastle disease vaccines. Biotechnol Adv. 1998:16:343–66.
- Webster Marketon JI, Glaser R. Stress hormones and immune function. Cell Immunol. 2008;252:16–26
- Puvadolpirod S, Thaxton J. Model of physiological stress in chickens 1. Response parameters. Poultry Sci. 2000;79:363–9.
- 5. Denbow DM, Snapir N, Furuse M. Inhibition of food intake by CRF in chickens. Physiol Behav. 1999;62:645–9.
- Sahin A, Forbes J. Corticosterone, dietary fat source and diet selection in relation to body composition. Brit Poultry Sci. 1999;40:52–4.
- Post J, Rebel J, Ter Huurne A. Physiological effects of elevated plasma corticosterone concentrations in broiler chickens. An alternative means by which to assess the physiological effects of stress. Poultry Sci. 2003:82:1313–8.
- Baxter J, Rousseau G. Glucocorticoid hormone action: An overview. Monogr Endocrinol. 1979;12:1–24.
- Siegel HS, Van Kampen M. Energy relationships in growing chickens given daily injections of corticosterone. Brit Poultry Sci. 1984;25:477–85.
- Dupont J, Tesseraud S, Simon J. Insulin signaling in chicken liver and muscle. Gen Comp Endocr. 2009;163:52–7.
- Li W, Yang X, Feng Y, Guo Y, Yao J. Effects of different immune stress on growth performance, nutrient digestibility and meat quality of broilers. Chinese J Anim Nutr. 2010;22:1551–7.
- Henken AM, Brandsma HA. The effect of environmental temperature on immune response and metabolism of the young chicken: 2. Effect of the immune response to sheep red blood cells on energy metabolism. Poultry Sci. 1982:61:1667–73.
- Siegel H, Henken A, Verstegen M, Van Der Hel W. Heat production during the induction of an immune response to sheep red blood cells in growing pullets. Poultry Sci. 1982;61:2296–300.
- Kang H, Wang H, Yu Q, Yang Q. A novel combined adjuvant strongly enhances mucosal and systemic immunity to low pathogenic avian influenza after oral immunization in ducks. Poultry Sci. 2013;92:1543–51.
- National Research Council. Nutrient requirements of poultry. 9th ed. Washington DC: National Academy Press; 1994.
- Zhang L, Yue H, Zhang H, Xu L, Wu S, Yan H, et al. Transport stress in broilers: I. Blood metabolism, glycolytic potential, and meat quality. Poultry Sci. 2009;88:2033–41.
- Zhang L, Guo Y. Effects of liquid DL-2-hydroxy-4-methylthio butanoic acid on growth performance and immune responses in broiler chickens. Poultry Sci. 2008;87:1370–6.
- 18. Association of Official Analytical Chemists. Official methods of analysis. 15th ed. Arlington, VA: Association of Official Analytical Chemists; 1990.
- Puvadolpirod S, Thaxton J. Model of physiological stress in chickens 4. Digestion and metabolism. Poultry Sci. 2000;79:383–90.
- Virden W, Lilburn M, Thaxton J, Corzo A, Hoehler D, Kidd M. The effect of corticosterone-induced stress on amino acid digestibility in ross broilers. Poultry Sci. 2007;86:338–42.
- Leeson S, Zubair A. Nutrition of the broiler chicken around the period of compensatory growth. Poultry Sci. 1997;76:992–9.

- Lippens M, Huyghebaert G, Groote GD. The efficiency of nitrogen retention during compensatory growth of food-restricted broilers. Brit Poultry Sci. 2002;43:669–76.
- 23. Yu M, Robinson F. The application of short-term feed restriction to broiler chicken production: A review. J Appl Poultry Res. 1992;1:147–53.
- Zubair A, Leeson S. Effect of early feed restriction and realimentation on heat production and changes in sizes of digestive organs of male broilers. Poultry Sci. 1994:73:529–38.
- 25. Siegel H. Stress, strains and resistance 1. Brit Poultry Sci. 1995;36:3–22.
- Virden W, Kidd M. Physiological stress in broilers: Ramifications on nutrient digestibility and responses. J Appl Poultry Res. 2009;18:338–47.
- Salak-Johnson J, McGlone J. Making sense of apparently conflicting data: Stress and immunity in swine and cattle. J Anim Sci. 2007;85:E81–8.
- Dhabhar FS. Enhancing versus suppressive effects of stress on immune function: Implications for immunoprotection and immunopathology. Neuroimmunomodulat. 2009;16:300–17.
- Korver D. Overview of the immune dynamics of the digestive system. J Appl Poultry Res. 2006;15:123–35.
- Leshchinsky T, Klasing K. Relationship between the level of dietary vitamin e and the immune response of broiler chickens. Poultry Sci. 2001;80:1590–9.

# Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at www.biomedcentral.com/submit

