

REVIEW

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Accurate models and nutritional strategies for specific oxidative stress factors: Does the dose matter in swine production?

Changming Hong^{1†}, Yujian Huang^{1†}, Shuting Cao¹, Li Wang¹, Xuefen Yang¹, Shenglan Hu¹, Kaiguo Gao¹, Zongyong Jiang¹ and Hao Xiao^{2*} 

Abstract

Oxidative stress has been associated with a number of physiological problems in swine, including reduced production efficiency. Recently, although there has been increased research into regulatory mechanisms and antioxidant strategies in relation to oxidative stress-induced pig production, it remains so far largely unsuccessful to develop accurate models and nutritional strategies for specific oxidative stress factors. Here, we discuss the dose and dose intensity of the causes of oxidative stress involving physiological, environmental and dietary factors, recent research models and the antioxidant strategies to provide theoretical guidance for future oxidative stress research in swine.

Keywords Accurate models, Dose, Nutritional strategies, Oxidative stress, Swine

Introduction

Oxidative stress has been interpreted as an imbalance between oxidation and anti-oxidation (more oxidizing), which was first defined in 1985 [1]. Oxidative stress in pigs is often associated with other pathological factors, including metabolic disorders and placental dysfunction in sows [2], and poor growth performance in piglets [2, 3]. These factors have a direct impact on sow reproductive performance and piglet growth. Therefore, it is crucial to address oxidative stress in pig production.

Current research efforts are primarily focused on alleviating oxidative stress in pigs by supplementing diets with antioxidants and free radical scavengers. However, the development of accurate models and nutritional strategies for specific oxidative stress factors has been largely unsuccessful.

The basis for the definition of oxidative stress is reactive oxygen species (ROS). ROS were the unregulated by-products of aerobic metabolism and other enzymatic processes that play a critical role in regulating cell function and biological processes. Uncontrolled production of ROS can overwhelm the ability of enzymatic and non-enzymatic antioxidant defence mechanisms, leading to a state of oxidative stress and consequently damage many biological macromolecules such as lipids, DNA and proteins [1]. The oxidative stress state can be measured, but find the specificity, has a strong correlation with physiological and pathological status of pigs accurately respond stress markers for oxidative stress measurement, specific display certain conditions, has prognostic value and structure stability of cost-effective, is particularly important to be used massively in pig production. In addition,

[†]Changming Hong and Yujian Huang contributed equally to this work.

*Correspondence:

Hao Xiao

xiaohao@gdaas.cn

¹ State Key Laboratory of Swine and Poultry Breeding Industry, Ministry of Agriculture Key Laboratory of Animal Nutrition and Feed Science in South China, Guangdong Public Laboratory of Animal Breeding and Nutrition, Guangdong Provincial Key Laboratory of Animal Breeding and Nutrition, Maoming Branch, Guangdong Laboratory for Lingnan Modern Agriculture, Guangzhou, China

² Institute of Animal Science, Guangdong Academy of Agricultural Sciences, 1 Dafeng 1st Street, Guangzhou 510640, China



dose ranges for indicators of whether pigs are under oxidative stress at different stages are also not yet well understood.

Here, we discuss the dose and dose intensity of the causes of oxidative stress including physiological status (pregnancy, lactation, neonatal or weaning stress), environmental factors (heat and cold stress) and dietary factors (dietary mycotoxins and lipid peroxidation) and regulatory metabolism in swine, the research models using different chemical compounds and also take into account the antioxidant strategies of nutritional regulation to provide theoretical guidance for the subsequent oxidative stress research.

Causes of oxidative stress in swine

Oxidative stress in swine can be caused by various factors, including physiological factors, environmental factors, and dietary factors. These causes have been the focus of recent studies on stress responses and regulatory metabolism in swine [4]. In this section, we focus on the levels of different stress indexes and effects at different physiological stages and aim to provide a comprehensive

understanding of the occurrence and mechanisms of oxidative stress in swine.

Physiologically induced oxidative stress

Oxidative stress in sows—gestation and lactation

It is well known that pregnant sows had increased oxidative stress during late gestation and lactation, which had an adverse effect on milk production, reproductive efficiency, and ultimately sow longevity [5]. Low DNA damage (21%) is present at d 30 of pregnancy (G 30), whereas increased DNA damage (38%–47%) is present throughout the gestational and lactational periods. Furthermore, plasma retinol and α -tocopherol concentration were reduced at the end of gestation (G 110) compared with G 30 [6]. Moreover, there is an increased systemic oxidative stress during late gestation and early lactation of sows, according to increasing levels of oxidative stress parameters, such as thiobarbituric acid reactive substances (TBARS), 8-hydroxy-2 deoxyguanosine (8-OHdG), and ROS [7] (Data are shown in Fig. 1). However, there is still lack studies focus on the reference value of oxidative stress in sows cause by gestation and lactation.

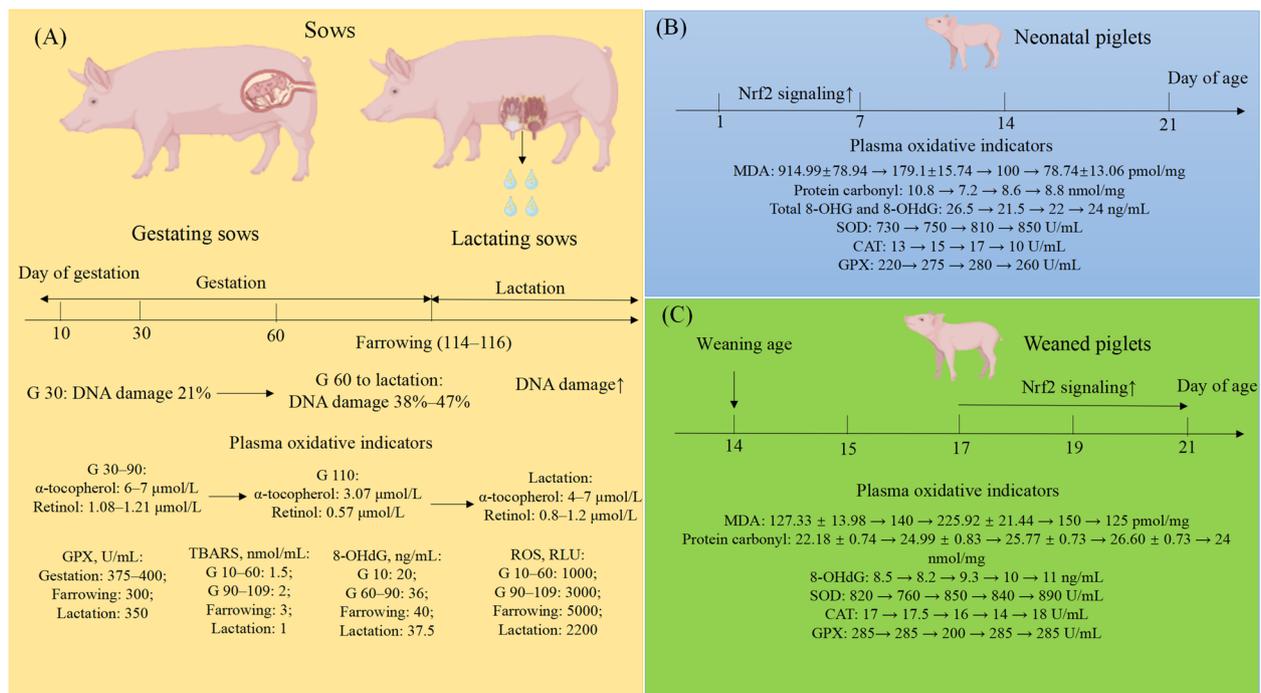


Fig. 1 Overview of physiologically induced oxidative stress in swine. **A** The increased DNA damage and oxidative stress parameters were exhibited at the end of gestation and lactation in sows. **B** For neonatal piglets, the highest oxidative stress parameters levels were exhibited at 1 d after birth of neonatal piglets. After that the oxidative stress parameters were reduced by the activation of Nrf2 signaling, which were reduced at 7 d after birth. **C** For weaned piglets, the highest oxidative stress parameters levels were exhibited at 3 d after weaning of 14 d weaned piglets. After that, the oxidative stress parameters were reduced by the activation of Nrf2 signaling. SOD: superoxide dismutase; Gpx: glutathione peroxidase; CAT: catalase; T-AOC: total antioxidant capacity; MDA: malondialdehyde; H_2O_2 : hydrogen peroxide; ROS: reactive oxygen species; 8-OHG: 8-hydroxyguanosine; 8-OHdG: 8-hydroxy-2 deoxyguanosine; TBARS: thiobarbituric acid reactive substances; RLU: relative light unit. The data in figure are come from the references [6–15]

Oxidative stress in piglets—birth and weaning stress

Due to their underdeveloped antioxidant systems, neonatal piglets are unable to efficiently scavenge excessive free radicals, resulting in oxidative stress [8]. Oxidative stress parameters like plasma malondialdehyde (MDA), protein carbonyl, and total 8-hydroxyguanosine (8-OHG) and 8-OHdG levels were exhibited at very high level at 1 d, while gradually decreased with increasing age. Furthermore, compared with the 1 d, superoxide dismutase (SOD) and glutathione peroxidase (GPx) activities were significantly increased ($P < 0.05$) at d 7, 14, and 21 [9] (Data are shown in Fig. 1). In addition, piglets with intra-uterine growth restriction (IUGR) exhibit oxidative stress characterized by lower antioxidant enzyme activities and increased lipid peroxidation in the liver compared to normal-birth piglets [10].

The effects of weaning stress on swine production have been well characterized [11–14], so this review will not describe them excessively. Generally, weaning stress could cause the changes in oxidative indicators that lead to oxidative stress in piglets (Fig. 1). For example, weaning induced oxidative stress was deteriorated until d 3 of weaning in 14 d weaned piglets. After that, the activation

of nuclear factor erythroid 2-related factor 2 (Nrf2) signaling may contribute to the alleviation of weaning stress [15]. Additionally, mitophagy, a process of selective degradation of damaged mitochondria, has been regarded to play a role in the feedback alleviation of oxidative stress after weaning [16]. Therefore, these pathways may be efficient targets for the nutritional regulation to alleviate weaning stress in piglets. Unfortunately, these articles mainly determine whether oxidative stress occurs according to the control group, as there is little research on the reference value of oxidative stress in weaning stress based on age and weight.

Environmentally induced oxidative stress

Pigs in particular are highly sensitive to changes in ambient temperature [17–19]. Therefore, this part focuses on how different environmental temperature-induced stresses of pigs, such as heat stress and cold stress, affect the performance of pigs at different stages. We also provide the thermoneutral zone for pigs at different stages with the aim of providing better regulatory strategies to alleviate environmentally induced oxidative stress in pigs (Fig. 2).

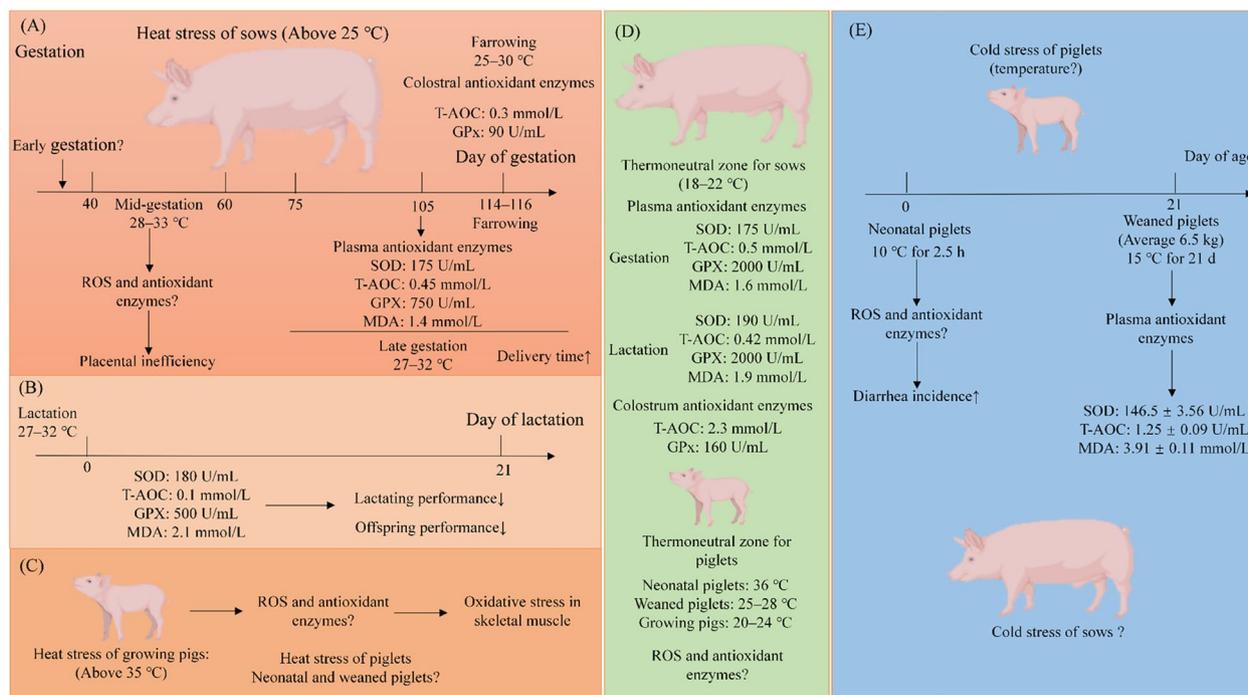


Fig. 2 Overview of environmentally induced oxidative stress in swine. **A** At mid gestation, heat stress causes the placental inefficiency of sows. At late gestation and farrowing, heat stress causes the decrease of antioxidant capacity, which prolong the delivery time of sows. **B** At lactation, the decrease of antioxidant capacity causes the decreased lactating performance of sows and further reducing the offspring performance. **C** Heat stress could lead to oxidative stress in skeletal muscle of growing piglets. **D** The thermoneutral zone and levels of oxidative indicators for pigs at different stage. **E** Cold stress could cause the increased diarrhea incidence of neonatal piglets. SOD: superoxide dismutase; Gpx: glutathione peroxidase; CAT: catalase; T-AOC: total antioxidant capacity; MDA: malondialdehyde; H₂O₂: hydrogen peroxide; ROS: reactive oxygen species. The data in figure are come from the references [20–31]

Heat stress

Sows that are metabolically stressed during the perinatal period are more susceptible to high temperatures, resulting in heat stress and reduced reproductive function [18]. The most suitable temperature for sows is 18–22 °C according to a previous study [20]. During mid-gestation (G 40–60), high summer temperatures (33 °C between 0900 and 1700 h, and 28 °C between 1700 and 0900 h) could lead to placental inefficiency in sows compared to a thermoneutral environment (constant 20 °C) [21]. During late gestation, compared to spring temperatures (from March to May, approximately 15–24 °C), high summer temperatures (from July to September, approximately 27.5–30 °C) reduced the antioxidant activities of plasma at G 105 (Gpx decreased from 2,000 to 750 U/mL) and colostrum at farrowing (Total antioxidant capacity (T-AOC) decreased from 2.3 to 0.3 mmol/L and Gpx decreased from 160 to 90 U/mL) in sows [22], which may prolong the parturition period of sows and result in poor growth performance of the offspring piglets [20, 23]. Around farrowing, sows exposed to high ambient temperatures (around 25–30 °C) also have an unfavourable effect on sow reproductive performance [24, 25]. Furthermore, at high summer temperatures (27–30 °C), lactating sows showed lower plasma antioxidant capacity (T-AOC decreased from 0.45 to 0.1 mmol/L and Gpx decreased from 2,000 to 500 U/mL) compared to those exposed to a thermoneutral environment (15–24 °C), which led to a decrease in lactation performance of sows, resulting in reduced performance of their offspring [22, 26]. For growing pigs, previous studies described that heat stress (35 °C for 1 or 3 d) could induce oxidative stress in skeletal muscle of growing pigs (35 ± 4 kg), possibly by inhibiting mitophagy and accumulation of damaged mitochondria in muscle cells [27, 28]. In addition, short-term heat stress (37 °C for 2–6 h) could induce oxidative stress in the muscle of growing pigs (63.8 ± 2.9 kg) [29]. These results suggest that heat stress has different effects at different stages in pigs. Unfortunately, there are few studies on the effects of heat stress on the performance of sows during early pregnancy and on neonatal and weaned piglets, and few studies focus on the value of oxidative stress in heat stress based on the stage of pigs, which needs further investigation.

Cold stress

Cold stress is another environmental challenge for swine, especially neonatal piglets. Previous research revealed that when exposed to a cold environment (10 °C) as opposed to a thermoneutral environment (36 °C), there was a decrease in the acquisition of colostrum immunoglobulin. This resulted in an increase in the incidence of diarrhea and may have contributed to preweaning

mortality in newborn piglets at 14.5 h of age [30]. Piglets exposed to a cold environment (15 °C) for 21 d showed higher antioxidant activities (T-AOC increased from 0.87 ± 0.05 to 1.25 ± 0.09 U/mL) than piglets exposed to a thermoneutral environment (26 °C), even though the animals' growth performance was diminished [31]. This may be because the piglets have become more adapted to prolonged cold exposure, as evidenced by the growth of oxidative muscle and gut bacteria [32–34]. Unfortunately, it is not yet known what temperature range causes cold stress in sows and weaned piglets. Furthermore, few studies have been conducted on the reference value of oxidative stress in piglets caused by cold stress. These should be prioritised in view of the rapid growth of the swine industry.

In conclusion, environmental challenges such as heat and cold stress may contribute significantly to oxidative stress in pig production. The possible thermoneutral zone and normal ranges of oxidative indicators for sows, neonatal and weaned piglets, and growing pigs are shown in Fig. 2.

Dietary-induced oxidative stress

Expect for physiologically and environmentally induced oxidative stress in swine, the dietary factors such as dietary mycotoxins pollution and lipid peroxidation could also cause oxidative stress in swine. In this section, the effects of different dose intensity of dietary mycotoxins and lipid peroxidation on the oxidative factors in various swine models will be summarized (Table 1).

Dietary mycotoxins-induced oxidative stress

The contamination of feed ingredients and complete feeds with these mycotoxins has been a major concern in China, especially in recent years [44]. Numerous studies have shown that the presence of these feed mycotoxins can lead to oxidative stress in pigs, reducing growth performance and meat quality [45–48]. In particular, the mycotoxin deoxynivalenol (DON) can induce oxidative stress and inflammation in the pig intestine, which has been extensively studied as a model of oxidative stress. Research has shown that supplementation of the basal diet of 28-day-old weaned piglets with 4 mg/kg DON resulted in a reduction in blood catalase (CAT) concentration [35]. Moreover, our recent studies have shown that dietary supplementation with 3.8 mg/kg DON reduced the intestinal antioxidant capacity and induced intestinal inflammation in 21 d weaned piglets [36, 37]. Dietary supplementation with 3–12 mg/kg DON could cause a decrease in antioxidant capacity in a dose-dependent manner, leading to oxidative stress in growing pigs [40, 41]. These results indicate that the peroxidative effects of DON on pigs are dependent on the dose of DON, the stage and the body weight of the pigs. In addition,

Table 1 Dietary-induced oxidative stress in swine

Swine models	Body weight, kg	Inducer and duration	Changes in oxidative indicators	References
Weaned piglets	12.36 ± 0.26	DON 4 mg/kg, 30 d (From 28 to 57 days of age)	Serum: D 15: H ₂ O ₂ (12 → 23 mmol/L) ↑ MDA (1 → 6.2 mmol/L) ↑ D 30: CAT (5.5 → 3.5 U/mL) ↓ H ₂ O ₂ (12 → 22 mmol/L) ↑ MDA (2 → 3 nmol/mL) ↑	[35]
	6.97 ± 0.1	DON 3.8 mg/kg, 28 d (From 21 to 49 days of age)	Serum: GSH (4.24 → 2.8 mg/L) ↓ T-AOC (2.31 → 1.56 U/mL) ↓ MDA (1.96 → 3.31 mmol/L) ↑ Ileum (μmol/g protein): GSH (5.13 → 4.21) ↓, SOD (58.24 → 39.57) ↓ T-AOC (0.49 → 0.31) ↓, MDA (0.52 → 1) ↑ Jejunum (μmol/g protein): GSH (10.93 → 10.02) ↓, SOD (63.66 → 49.95) ↓ T-AOC (0.4 → 0.29) ↓, MDA (0.61 → 0.8) ↑	[36, 37]
	10.9 ± 0.77	OTA 0.25 mg/kg, 28 d	Liver: T-AOC (27 → 22 μmol/g tissue) ↓ TBARS (3.5 → 5.4 nmol/g tissue) ↑ Kidney: GPx (1.8 → 22 μmol/min/g tissue) ↓ T-AOC (23 → 19 μmol/g tissue) ↓ TBARS (0.8 → 1.2 nmol/g tissue) ↑	[38]
	6.97 ± 0.1	AFB1 0.32 mg/kg, 30 d (From 28 to 57 days of age)	MLNs: SOD (400 → 352 U/g tissue) ↓ CAT (4 → 2.92 μmol/min/g tissue) ↓ GPx (4.8 → 3.74 μmol/min/g tissue) ↓ T-AOC (11 → 8.75 μmol/g tissue) ↓	[39]
Growing pigs	16.3 ± 1.5	DON 3–12 mg/kg, 21 d (From 67 to 88 days of age)	Serum (U/mL): 3 mg/kg: SOD (117.5 ± 6.26 → 100.88 ± 14.24) ↓ 6 mg/kg: SOD (117.5 ± 6.26 → 84.48 ± 8.97) ↓ GPx (3,424.7 ± 147.5 → 2,967.3 ± 124.4) ↓ 12 mg/kg: SOD (117.5 ± 6.26 → 79.94 ± 15.62) ↓ GPx (3,424.7 ± 147.5 → 2,521.3 ± 334.6) ↓	[40, 41]
Nursery pigs	Average 6.5	Oxidized soybean oil 60 g/kg, 35 d (From 21 to 66 days of age)	Jejunal mucosa: MDA (35 → 54 μmol/g protein) ↑	[42]
Newborn piglets	1.8 ± 0.04	Oxidized soybean oil 50 g/kg, 21 d (From 4 to 25 days of age)	Jejunal mucosa: MDA (3.17 → 5.2 mol/L) ↑ GSSG (8.58 → 12.62 mmol/g protein) ↑	[43]

CAT Catalase, GPx Glutathione peroxidase, SOD Superoxide dismutase, MLNs Mesenteric lymph nodes, DON Deoxynivalenol, OTA Ochratoxin A, AFB1 Aflatoxin B1, ZEA Zearalenone

exposure of piglets to 0.25 mg/kg ochratoxin A (OTA) reduced antioxidant capacity in the liver and kidney [38]. Moreover, dietary supplementation with 0.32 mg/kg aflatoxin B1 (AFB1) decreased the antioxidant status in piglet mesenteric lymph nodes (MLNs) [39], which may contribute to intestinal barrier dysfunction [49]. However, future studies should focus on oxidative stress caused by dietary mycotoxins based on dosage and stage, age or weight of the pigs.

Dietary lipid peroxidative induced stress

Lipids are susceptible to be oxidized at high temperatures and produce large amounts of lipid peroxides (e.g., MDA and 4-hydroxynonenal (4-HNE)), which could lead to lipid peroxidation in animals [50, 51]. Therefore, proper assessment of lipid peroxidation has been an important part of quality control for the prevention of oxidative stress and performance loss in pigs

[52]. Dietary supplementation with 60 g/kg oxidized soybean oil (MDA level = 4.5 mmol/L oil) increased lipid peroxidation (characterized by MDA levels ranging from 35 to 54 μmol/g protein in the jejunal mucosa) of nursery pigs [42]. Our previous studies demonstrated that dietary supplementation with 50 g/kg oxidized fish oil (peroxide level = 186.89 mmol/kg) significantly increased the levels of MDA and glutathione oxidized (GSSG) in the jejunal mucosa of newborn piglets compared to those dietary added 50 g/kg fresh fish oil (peroxide level = 4.20 mmol/kg) [43]. Cellular models of 4-HNE-induced oxidative stress have also been used to study the peroxidative effects of a lipid peroxidation diet on pigs [53, 54]. These studies found that 4-HNE could induce an increase in cellular ROS generation and DNA damage in IPEC-1 cells [55, 56]. However, lipid peroxidation in swine diets has not yet been well studied.

Table 2 Chemical compounds-induced oxidative stress models in swine

Swine models	Body weight, kg	Inducer and duration	Changes in oxidative indicators	References
Weaned piglets	9.92 ± 0.3	Diquat 10 mg/kg, 7 d	Serum: T-AOC (2.5 → 2.1 U/L) ↓ MDA (2.8 → 4.5 nmol/L) ↑	[57]
	10.63 ± 1	Diquat 10 mg/kg, 7 d (From 35 to 42 days of age)	Serum: SOD (74.8 ± 1.5 → 56.14 ± 1.58 U/mL) ↓ CAT (10.35 ± 0.53 → 8.97 ± 0.34 U/mL) ↓ GPx (493.62 ± 30.95 → 313.56 ± 48.67 U/mL) ↓ MDA (2.76 ± 0.07 → 3.32 ± 0.11 mmol/L) ↑	[58]
	Average 9.6	Diquat 10 mg/kg, 7 d (From 35 to 42 days of age)	Jejunum: SOD (100.32 ± 11.29 → 60.06 ± 8.01 U/mg protein) ↓ GPx (99.35 ± 8.31 → 57.88 ± 6.4 U/mg protein) ↓ MDA (0.54 ± 0.07 → 1.91 ± 0.1 nmol/g protein) ↑ Mitochondrial ROS (9 times higher than control) ↑	[59]
	10.96 ± 0.61	H ₂ O ₂ 1 mg/kg, 7 d	Serum: MDA (3.8 → 4.5 ng/mL) ↑ H ₂ O ₂ (20 → 28 mmol/L) ↓	[60]
	11.58 ± 0.26	LPS 100 µg/kg, 3 h (On 35 days of age)	Serum: SOD (110.6 ± 6.9 → 98.3 ± 5.8 U/mL) ↓ CAT (4.75 ± 0.51 → 4.1 ± 0.36 U/mL) ↓ GPx (396.6 ± 18.5 → 359.4 ± 36.8 U/mL) ↓ H ₂ O ₂ (51.28 ± 8.09 → 65.05 ± 8.46 mol/L) ↑ MDA (5.63 ± 0.46 → 6.59 ± 1 mmol/L) ↑	[61]
	Average 9.8	LPS 100 µg/kg, 7 d (From 35 to 42 days of age)	Jejunum: SOD (95.85 ± 3.13 → 63.12 ± 2.97 U/mg protein) ↓ GPx (72.11 ± 3.51 → 53.99 ± 3.26 U/mg protein) ↓ MDA (0.95 ± 0.16 → 1.51 ± 0.25 nmol/g protein) ↑ Mitochondrial ROS (8 times higher than control) ↑	[62]
	6.65 ± 1.19	LPS 100 µg/kg, 5 d (From 31 to 36 days of age)	Serum: T-AOC (0.21 ± 0.008 → 0.16 ± 0.025 mmol/L) ↓ SOD (27.73 ± 1.19 → 24.73 ± 1.59 U/mL) ↓ GPx (317.2 ± 28.96 → 245.63 ± 21.37 U/mL) ↓	[63]
100-day-old male miniature pigs	21.73 ± 0.43	CTX 50 mg/kg, 7 d	Serum: SOD (27 → 8 U/L) ↓ GPx (420 → 220 µmol/L) ↓ MDA (12 → 1.51 ± 24 µmol/L) ↑	[64]

BW Body weight, *CAT* Catalase, *SOD* Superoxide dismutase, *GPx* Glutathione peroxidase, *MDA* Malondialdehyde, *ROS* Reactive oxygen species, *H₂O₂* Hydrogen peroxide, *GSH* Glutathione, *T-AOC* Total antioxidant capacity, *LPS* Lipopolysaccharide

Chemical compounds-induced oxidative stress models in swine

Many chemical compounds (e.g., diquat, hydrogen peroxide (H₂O₂) and lipopolysaccharide (LPS)) have also been used to investigate the underlying mechanism of oxidative stress in swine. Here, we will discuss the regulatory mechanisms of each of the oxidative stress models, and then compare their respective characteristics and assess their impact on the pig mode (Table 2).

Diquat-induced oxidative stress models

Diquat is a herbicide that is commonly used to induce oxidative stress in animal models due to its duration [65], making it a suitable model to study how oxidative stress affects physiological changes in swine [66]. Injection of 10 mg/kg diquat for 7 d reduced T-AOC levels and increased MDA levels, causing intestinal damage and reducing growth performance in weaned piglets [57]. Furthermore, 35-day-old weaned piglets treated with

10 mg/kg diquat for 7 d showed a significant decrease of antioxidant capacity and increased MDA levels [58], which could lead to mitochondrial dysfunction and further impairment of intestinal barrier function [59]. Eventually, diquat leads to an excess of oxygen radicals and aminotransferase in pigs, which disturbs the redox balance in the liver or gut. In conclusion, models of diquat-induced oxidative stress are focused on weaned piglets between 9–10 kg, and the dose of diquat is 10 mg/kg (Table 2).

H₂O₂-induced oxidative stress models

H₂O₂-induced oxidative stress in porcine intestine epithelium cells, as shown by elevated ROS levels, resulting in reduced cell viability [67]. Intragastric or intraperitoneal treatment with 100 µmol/L H₂O₂ caused oxidative stress in intestinal epithelial cells through the blockade of T-AOC [68]. 1 mg/kg H₂O₂ increased serum MDA and H₂O₂ levels in piglets by activation of the nuclear

factor- κ B (NF- κ B) and Nrf2 signaling pathways and induction of the autophagic response in the jejunum [60, 69]. Furthermore, high doses H₂O₂ (10 mg/kg for piglet model (11.29 ± 0.32 kg) or 100 μmol/L for cell model) promoted mitochondrial dysfunction, whereas low doses H₂O₂ (5 mg/kg for piglet model or 50 μmol/L for cell model) showed a feedback regulatory mechanism against mitochondrial oxidative damage by increasing uncoupling protein 2 (UCP2) expression and mitochondrial proton leak [70]. Mitophagy may be involved in the above feedback control to improve oxidative stress (induced by 600 μmol/L H₂O₂)-induced intestinal barrier degradation [71]. In conclusion, H₂O₂ is the direct inducer of cellular oxidative stress. However, the H₂O₂-induced oxidative stress model has significant limitations in its application to pigs because pigs must first undergo surgery, which can easily lead to gastrointestinal ulceration in pigs, reducing their survival rates.

LPS-induced oxidative stress models

LPS, a lipid and polysaccharide endotoxin, has been used to induce an inflammatory response in most animal studies [72]. However, because of the link between the inflammatory response and oxidative stress [73], the LPS model has also been used as a model of oxidative stress in porcine research. For example, LPS at 100 mg/kg (intraperitoneal injection) caused intestinal oxidative stress in weaned piglets, as indicated by decreased jejunal T-AOC and GSH levels and increased MDA levels [74]. Piglets exposed to LPS also showed impaired intestinal barrier and mitochondrial function, as well as facilitated intestinal mitophagy [62]. In addition, after continuous low-dose induction of LPS, weaned piglets may have developed tolerance to endotoxin. LPS-induced oxidative stress was observed at an early stage (within 5 d of LPS stimulation), but was attenuated at a later stage (after 5 days of LPS stimulation) [63]. Overall, LPS induces oxidative stress in animals, mostly through the induction of inflammatory responses. Therefore, the LPS-induced oxidative stress paradigm is excellent for studying nutrients that have both antioxidant and anti-inflammatory properties, although individual studies of oxidative stress are less effective.

Other chemical compounds induced oxidative stress models

Other chemicals (including D-galactose and cyclophosphamide (CTX)) have been used in pigs. In IPEC-J2 cells, treatment with 500 μmol/L CTX resulted in a significant decrease in the activity of antioxidant enzymes and a significant increase in the MDA level [75]. Furthermore, dietary treatment with 50 mg/kg CTX dramatically decreased antioxidant enzyme activities while significantly increasing MDA levels [64]. Dietary D-galactose at 10 g/kg BW significantly increased blood MDA

levels while decreasing intestinal antioxidant capacity in weaned piglets [76]. Although few studies have been conducted in porcine models, these findings suggest that CTX and D-galactose may be potential inducers of oxidative stress. In addition, the specific doses of the chemical compounds in pigs at different stages of oxidative stress should also be taken into consideration.

Nutritional regulation strategies for oxidative stress in swine production

Recently a number of nutrients have been in use for the reduction of oxidative stress in pig production [77–79]. In this section, we mainly review current research on polyphenols and functional amino acids to reduce oxidative stress in pig production.

Polyphenols

Polyphenols, the secondary plant metabolites with antioxidant properties, are attractive feed additives for nutritional management in pig production [80, 81]. In this section, we review current research on polyphenols for the reduction of oxidative stress in pigs (Table 3).

Daidzein

Daidzein is commonly used in pigs for its antioxidant properties [98]. Dietary supplementation with 50 mg/kg daidzein improved growth performance, increased SOD activity and decreased plasma MDA in weaning pigs [82]. Dietary supplementation with a high dose of daidzein (640 mg/kg) increased the antioxidant capacity of the longissimus muscle, but had a pro-oxidant effect on the back fat, abdominal fat, liver and plasma of finishing pigs [83]. Moreover, 40 mol/L of daidzein increased the expression of the Nrf2, CAT, and occludin genes in H₂O₂-stimulated IPEC-J2 cells [82], suggesting that dietary daidzein may have a beneficial role in the health of the intestine in pigs.

Daidzein is also a type of active phytoestrogen that is beneficial to reproductive performance in sows. In our previous studies, dietary supplementation with 40 mg/kg daidzein from G 75 to L 21 markedly increased the activities of antioxidant enzymes in sow colostrum, while had no effect on serum antioxidant capacity in pregnant sows [84]. Notably, dietary supplementation with 200 mg/kg daidzein may increase serum antioxidant capacity in pregnant sows [85]. We speculated that daidzein is needed more during pregnancy than during lactation to support placenta development.

Resveratrol and its derivatives

Resveratrol (3,4',5-trihydroxystilbene) is known for its antioxidant properties in livestock, particularly pigs [99]. For example, dietary resveratrol at 90 mg/kg effectively reduced diquat-induced oxidative stress in piglets

Table 3 The antioxidant effects of polyphenols on swine

Polyphenols	Dosages and duration	Swine models	Antioxidant effects	References
Daidzein	50 mg/kg 72 d	Weaned piglets 23-day-old 7.35 ± 0.14 kg BW	Plasma: D 14: MDA (2.52 → 2.31 mmol/L) ↓ D 42: SOD (18.19 → 20.54 U/mL) ↑	[82]
	640 mg/kg 64 d	Finishing pigs Average 57 kg BW	Liver (U/mg protein): CAT (79.5 → 96.3) ↑, SOD (285 → 358) ↑ longissimus muscle (U/mg protein): T-AOC (0.036 → 0.063) ↑, SOD (14.71 → 17.56) ↑	[83]
	40 mg/kg G75 to L21	Sows	Colostrum (U/mL): SOD (119.2 ± 4.14 → 154.8 ± 8.99) ↑ CAT (2.4 ± 0.58 → 18.73 ± 1.1) ↑ GPx (155.3 ± 9.76 → 201.4 ± 18.73) ↑ T-AOC (2.15 ± 0.15 → 2.72 ± 0.12) ↑	[84]
	200 mg/kg G1 to G114	Sows (3 to 5 parity, 226.18 ± 1.09 kg BW on G35 239.64 ± 0.83 kg BW on G85)	Serum (U/mL): G35: GPx (1,169.9 → 1,486.8) ↑ SOD (94.39 → 99.92) ↑; CAT (12.21 → 14.5) ↑ T-AOC (6.03 → 7.12) ↑ G85: SOD (62.28 → 69.58) ↑, T-AOC (5.53 → 7.7) ↑	[85]
Resveratrol	90 mg/kg 21 d	Diquat-challenged weaned piglets 28-day-old 7.25 ± 0.13 kg BW	Jejunal mucosa (U/mg protein): CAT (5.56 → 6.68) ↑, SOD (50.33 → 74.48) ↑ GPx (46.25 → 59.64) ↑ MDA (0.96 → 0.5 nmol/g protein) ↓	[86]
	100 mg/kg 14 d	Diquat-challenged weaned piglets 35-day-old 9.35 ± 0.26 kg BW	Jejunum: T-AOC (0.18 → 0.33 U/mg protein) ↑ H ₂ O ₂ (20.43 → 15.61 mmol/g protein) ↓ MDA (1.05 → 0.59 nmol/mg protein) ↓	[87]
	300 mg/kg 28 d	DON-challenged weaned piglets 21-day-old 6.97 ± 0.1 BW	Serum: GSH (2.8 → 3.38 mg/L) ↑ T-AOC (1.56 → 2.01 mg/L) ↑ MDA (3.31 → 2.58 nmol/mL) ↓ Ileum (μmol/g protein): SOD (39.57 → 50.77) ↑, T-AOC (0.31 → 0.44) ↑, MDA (1 → 0.68) ↓ Jejunum (μmol/g protein): GSH (10.02 → 11.17) ↑, SOD (49.95 → 57.76) ↑ T-AOC (0.29 → 0.37) ↑	[36, 37]
		Oxidized soybean oil challenged piglets Average 34.43-day-old 10.19 ± 0.1 kg BW	Plasma H ₂ O ₂ (82.99 → 70.11 mmol/L) ↓	[88]
	300 mg/kg G75 to L21	Sows at high temperatures (average parity 3)	Plasma: L14: T-AOC (0.1 → 0.15 mmol/L) ↑ MDA (2.1 → 1.6 nmol/mL) ↓ Colostrum: MDA (3.8 → 2 nmol/mL) ↓	[22]
300 mg/kg G20 to L21	Sows (average parity 4.4)	Plasma: G110: CAT (12 → 14 U/mL) ↑, GPx (2,200 → 2,800 U/mL) ↑ MDA (2.5 → 1.5 nmol/mL) ↓, H ₂ O ₂ (50 → 38 mmol/L) ↓ L14: SOD (80 → 120 U/mL) ↑; CAT (2.5 → 4 U/mL) ↑ MDA (1.6 → 1.2 nmol/mL) ↓, H ₂ O ₂ (18 → 15 mmol/L) ↓ L21: SOD (80 → 120 U/mL) ↑; MDA (1.7 → 1.4 nmol/mL) ↓ Placenta (U/mg protein): CAT (28 → 40) ↑, SOD (28 → 48) ↑, GPx (75 → 90) ↑, MDA (5.8 → 3) ↓, H ₂ O ₂ (18 → 15 mmol/g protein) ↓	[89]	
Pterostilbene	300 mg/kg 15 d	DON-challenged weaned piglets 21-day-old 6.57 ± 0.32 kg BW	Liver: MnSOD (25.5 ± 4.96 → 53.75 ± 5.39 U/mg protein) ↑ GSH (63.11 ± 6.95 → 88.78 ± 6.34 nmol/100 mg wet weight) ↑	[90]
	300 mg/kg 14 d	IUGR piglets 21-day-old 4.91 ± 0.06 kg BW	Jejunum: SOD (60 → 90 U/mg protein) ↑ GSH (1.8 → 2.8 μmol/100 mg wet weight) ↑ T-GSH (3 → 4 μmol/100 mg wet weight) ↑ MDA (0.6 → 0.45 nmol/mg protein) ↓	[91]

Table 3 (continued)

Polyphenols	Dosages and duration	Swine models	Antioxidant effects	References
Curcumin	400 mg/kg 24 d	IUGR piglets 26-day-old	Serum: T-AOC (1.03 ± 0.37 → 1.89 ± 0.57 U/mL) ↑ CAT (1.77 ± 0.43 → 3.59 ± 1.27 U/mL) ↑ GPx (355.29 ± 50.57 → 376.87 ± 25.39 U/mL) ↑ GR (21.47 ± 1.38 → 30.14 ± 1.24 U/L) ↑ Liver: T-AOC (0.88 ± 0.14 → 1.21 ± 0.22 U/mg protein) ↑ GPx (36.23 ± 2.04 → 58.30 ± 5.17 U/mg protein) ↑ GR (16.11 ± 1.22 → 18.94 ± 3.36 U/g protein) ↑	[92]
	200 mg/kg 89 d	IUGR piglets 26-day-old 5.76–6.05 kg BW	Jejunum: SOD (15.82 ± 3.04 → 28.65 ± 4.83 U/mg protein) ↑ MDA (2.01 ± 0.15 → 1.23 ± 0.13 nmol/mg protein) ↓ Leg muscle (U/mg protein): SOD (16.69 ± 0.23 → 17.83 ± 0.28) ↑ GPx (13.65 ± 0.53 → 15.50 ± 0.82) ↑ CAT (14.18 ± 0.32 → 18.21 ± 0.97) ↑ MDA (4.65 ± 0.32 → 3.42 ± 0.19 nmol/mg protein) ↓	[93, 94]
Polyphenols	Dosages and duration	Swine models	Antioxidant effects	References
Curcumin	200 mg/kg 169 d	IUGR piglets 26-day-old	<i>longissimus dorsi</i> muscle GSH (27.99 ± 2.21 → 40.43 ± 1.80) ↑ CAT (10.56 ± 0.58 → 12.59 ± 0.83) ↑ T-AOC (0.32 ± 0.04 → 1.22 ± 0.38) ↑ MDA (2.57 ± 0.17 → 1.08 ± 0.08 nmol/mg protein) ↓	[95]
Eucommia ulmoides flavones	100 mg/kg 21-d feeding	DON-challenged weaned piglets 21-day-old 6.50 ± 0.29 kg BW	Serum (U/mL): D 14: SOD (73.51 ± 6.31 → 94.87 ± 3.37) ↑ GPx (335.21 ± 4.14 → 360.85 ± 2.49) ↑ CAT (5.73 ± 0.38 → 6.97 ± 0.38) ↑ T-AOC (0.54 ± 0.01 → 1.14 ± 0.1) ↑ D 21: GPx (314.49 ± 5.68 → 334.15 ± 4.41) ↑	[96]
	100 mg/kg 14-d feeding		Jejunum: SOD (7.678 ± 2.068 → 13.263 ± 1.67 U/mg protein) ↑ MDA (1.698 ± 0.164 → 3.356 ± 0.546 nmol/mg protein) ↓	[97]

BW Body weight, SOD Superoxide dismutase, Gpx Glutathione peroxidase, CAT Catalase, T-AOC Total antioxidant capacity, MDA Malondialdehyde, H₂O₂ Hydrogen peroxide, GSH Glutathione, GR Glutathione reductase

through activation of Nrf2 pathways [86]. Resveratrol (15 mol/L) was also found to reduce intracellular ROS accumulation and increase cell viability in IPEC-J2 cells through activation of the Nrf2 pathway [100]. Both in vivo and in vitro experiments show that resveratrol modulates mitophagy in DON-injured piglets [101]. Dietary resveratrol (100 mg/kg) increased intestinal redox status by inducing intestinal mitophagy in diquat-challenged piglets [87]. Furthermore, dietary resveratrol (300 mg/kg) can also ameliorate oxidative stress induced by oxidized soybean oil [88] or DON [36, 37] by altering the intestinal microbiota. These results suggest that different doses of resveratrol may alleviate oxidative stress in weaned piglets through different pathways.

Previous research found that dietary 300 mg/kg resveratrol during pregnancy and lactation increased antioxidant capacity in sows [89]. Similarly, our recent studies found that maternal resveratrol at 300 mg/kg during late gestation and lactation significantly increased sow plasma T-AOC from lactation and decreased sow plasma and colostrum MDA at high summer temperatures,

influenced by gut microbiota [22]. However, there is still a lack of studies focusing on the antioxidant effects of other doses of resveratrol in sows, which is in need of further investigation.

Pterostilbene is a methylated derivative of resveratrol, which has recently been used in pig production because of its superior antioxidant effects compared to resveratrol [90, 91, 102]. However, while resveratrol has been under investigation in pig production for almost 10 years [103], pterostilbene has only been under investigation for 3 years [90]. Further studies are needed to investigate the antioxidant effects of pterostilbene and other resveratrol derivatives in pig production.

Curcumin

Curcumin, a natural lipophilic polyphenol derived from the turmeric rootstock, has been shown to possess antioxidant properties in swine [104]. Dietary supplementation with 400 mg/kg curcumin increased hepatic antioxidant capacity by increasing the expression of Nrf2 and Hmox1, resulting in improved growth performance

in weaned piglets with IUGR [92]. Similarly, dietary curcumin supplementation at 200 mg/kg can improve redox status by activating the Nrf2 signaling pathway in leg muscles, *longissimus dorsi* and jejunum [93–95], which may improve the meat quality or reduce intestinal damage respectively in growing IUGR pigs. Curcumin significantly reduced oxidative stress-induced intestinal damage and mitochondrial dysfunction in piglets by promoting Parkin-dependent mitophagy [105]. Moreover, curcumin reduced diquat-induced intestinal oxidative damage and mitochondrial dysfunction by reducing endoplasmic reticulum stress and inhibiting apoptosis [106]. These results suggest that curcumin alleviates oxidative stress in pigs by ameliorating mitochondrial dysfunction, endoplasmic reticulum stress and apoptosis, and by activating the Nrf2 signaling pathway. The recommended dose of curcumin for pigs is (200 to 400 mg/kg body weight).

Eucommia ulmoides flavones

Eucommia ulmoides flavones (EUFs) are secondary metabolites from *Eucommia ulmoides* (a Chinese herb with various medicinal properties [107]) which have recently been shown to have potent antioxidant activities in pigs. Dietary supplementation with 100 mg/kg EUF attenuated diquat-induced oxidative stress, inflammatory response and impaired growth in piglets [96]. The Nrf2 signaling pathway has been shown to play a critical role in EUF-mediated alleviation of intestinal oxidative stress in diquat-treated piglets. [97]. These findings may have implications for the investigation of EUFs as potential antioxidants in pig production. However, further studies are needed to investigate whether and how EUFs exert antioxidant effects in pigs at other stages of life, such as in finishing pigs and in perinatal sows.

Taken together, dietary supplementation with polyphenols may have good antioxidant effects in pigs. However, the underlying mechanism by which polyphenols exert antioxidant effects is in need of further investigation.

Functional amino acids

Functional amino acids and their bioactive precursors such as nitric oxide (NO), polyamines and GSH could reduce oxidative stress and improve pig growth, reproduction and health (Fig. 3) [108, 109]. For example, supplementation with 1% arginine dramatically increased antioxidant capacity in serum and skeletal muscle to improve meat quality in growing pigs [110]. Dietary supplementation with 0.8% and 1.6% arginine significantly increased plasma and liver GPx and SOD activities in diquat-challenged weaned piglets to alleviate oxidative stress responses [111]. It is interesting to note that arginine could be metabolized to NO to alleviate

oxidative stress in sows [112–114]. Furthermore, our previous studies showed that arginine administration significantly increased GPx activity and reduced ROS and MDA production via the arginase-1 pathway in LPS-stimulated IPEC-J2 cells [115].

Cysteine, glutamate and glycine are precursors of GSH synthesis. Therefore, these three amino acids may influence antioxidant function [116]. Cysteine supplementation increased intestinal CAT, SOD, GPx and GSH activities for maintenance of intestinal integrity in LPS-challenged weaned piglets [117]. Supplementation with 1.2 g/kg *N*-acetylcysteine (NAC) (a precursor of cysteine) increased GSH levels and decreased MDA levels in the liver of IUGR suckling piglets [118]. Dietary supplementation with 500 mg/kg NAC increased liver antioxidant capacity in weanling piglets to attenuate LPS-induced liver injury [61]. Treatment with 800 μ mol/L NAC increased the antioxidant capacity to reduce cell apoptosis in H₂O₂-induced IPEC-J2 cells [68], suggesting that NAC could rescue H₂O₂-induced intestinal oxidative damage in piglets. Moreover, treatment with 5 mmol/L NAC attenuated 4-HNE-induced cell death in intestinal epithelial cells through activating the Nrf2 signaling pathway [53]. Furthermore, increased supplementation of cysteine in the diet (from 0.3% to 0.4% and 0.5%) could reduce plasma MDA levels to reduce oxidative stress in sows during late gestation and lactation [119]. Dietary NAC supplementation (500 mg/kg) significantly increased serum and placental antioxidant capacity (e.g., GSH levels) and activated the STAT3/occludin/ZO-1 pathway in the placentas of sows to attenuate DON-induced placental oxidative stress and barrier damage, further reducing the incidence of stillbirths and low birth weight piglets [120].

Glutamate also possesses antioxidant properties [121]. Dietary supplementation with 2% glutamate increased plasma SOD and GPx activity and promoted intestinal epithelial cell proliferation in DON-challenged piglets [122]. Furthermore, glutamate supplementation reduced MDA production and ameliorated diquat-induced oxidative stress in piglets by increasing SOD, T-AOC and NO levels while decreasing MDA production [57]. Furthermore, dietary supplementation with glutamine (a precursor of glutamate) can increase intestinal antioxidant capacity [123] and the expression of genes involved in GSH synthesis (e.g., glutamate cysteine ligase catalytic (*GCLC*), glutamate cysteine ligase modifier (*GCLM*), and glutathione reductase (*GSR*)) and 4-HNE metabolism (e.g., glutathione S-transferase A 1 (*GSTA1*) and *GSTA4*) to attenuate 4-HNE-induced oxidative stress [55].

Glycine, one of the key components of GSH, is of great importance in the treatment of oxidative stress in

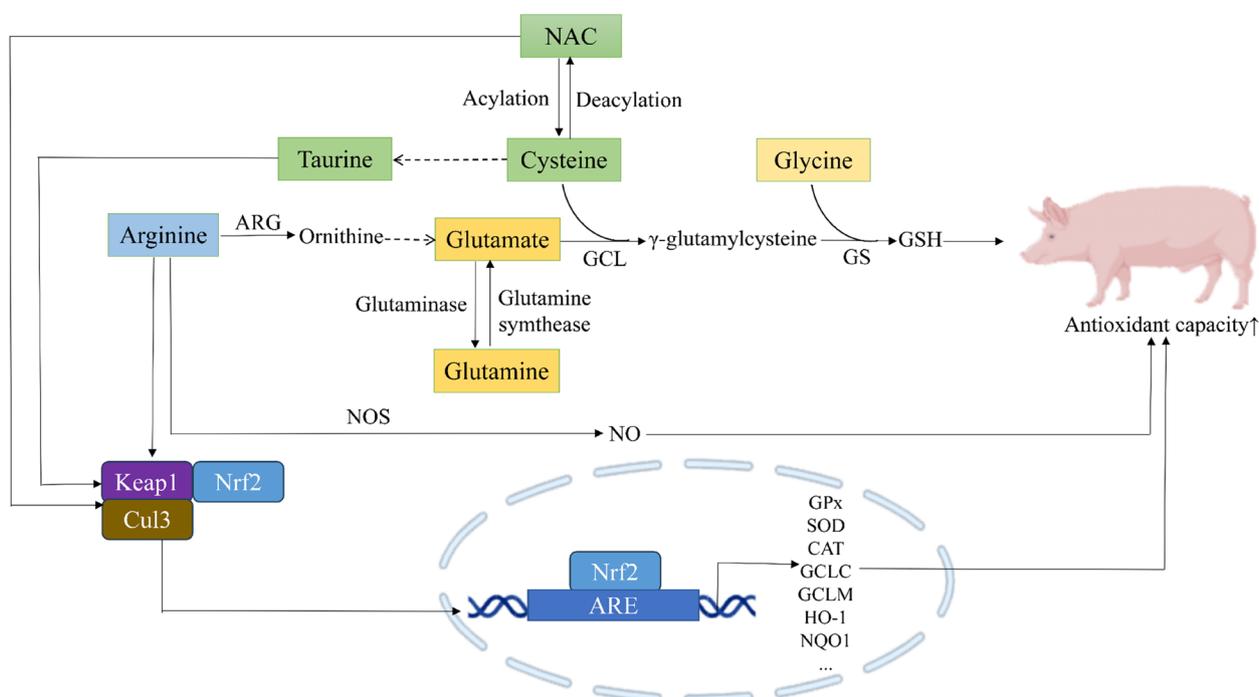


Fig. 3 The proposed molecular mechanisms by which functional amino acids improve the antioxidant capacity of swine. Arginine supplementation in swine can be catalyzed to ornithine, and then transfer into glutamate, which conjugates cysteine to generate γ -glutamyl cysteine that finally conjugates glycine to form GSH to improve the antioxidant capacity. Supplemented with glutamate, cysteine and glycine, or its precursors glutamine and NAC in swine can also improve the antioxidant capacity via GSH synthesis. Arginine, taurine and NAC can perform antioxidant effects through the activation of Nrf2 pathway, and arginine can also improve antioxidant capacity of swine via NO synthesis stimulated by NOS. NAC: *N*-acetylcysteine; ARG: arginase; NOS: oxide synthase; NO: nitric oxide; GS: synthase; GSH: glutathione; Nrf2: nuclear factor erythroid 2 (NF-E2)-related factor 2; Keap1: Kelch-like ECH-associated protein 1; Cul3: Cullin3; ARE: antioxidant responsive element; GPx: glutathione peroxidase; SOD: superoxide dismutase; CAT: catalase; GCLC: glutamate cysteine ligase catalytic; GCLM: glutamate cysteine ligase modifier; HO-1: heme oxygenase; NQO1: NAD(P)H:quinone oxidoreductase 1

pigs. Dietary 0.5%–2% glycine supplementation linearly increased plasma and intestinal glycine and GSH levels in milk-fed piglets [124]. Dietary supplementation with 1% glycine improved intestinal mucosal morphology and antioxidant capacity (e.g., GSH and GPx activities), and inhibited the ferroptosis through regulating key proteins (e.g., transferrin receptor 1, SLC7A11 and GPx4) in intestinal mucosa of diquat-challenged weanling piglets [125]. Furthermore, treatment with 1 mmol/L glycine significantly increased GSH activity, protein synthesis and cell proliferation, and decreased cell apoptosis in the 4-HNE-induced oxidative stress model of IPEC-1 cells [126]. These results suggest that glycine may inhibit lipid peroxidation and intestinal epithelial ferroptosis by reducing oxidative stress in piglets through the production of GSH and GPx.

Taurine is an amino acid derived from converting sulphur-containing amino acids. Dietary taurine levels of 0.3% to 0.6% increased the activity of antioxidant enzymes (e.g., T-SOD and GPx) and reduced the levels of oxidative indicators (ROS, 8-OHdG and MDA) by increasing and activating the Nrf2 signaling system

in weaned piglets [127]. Besides, dietary supplementation with 1% taurine from G 75 to lactation significantly increased the antioxidant capacity in the gilts' milk and the intestinal antioxidant effects in the piglets [128]. Furthermore, pre-treatment with 2 mmol/L taurine reduced H₂O₂-induced oxidative stress in PMECs by increasing SOD activity and decreasing intracellular ROS [129]. These results suggest that dietary taurine supplementation may improve antioxidant capacity and milk performance via the Nrf2 pathway in lactating sows.

In conclusion, dietary supplementation with functional amino acids has potential antioxidant effects in pigs, but this requires further investigation using different models of oxidative stress, functional amino acids, and dietary levels or proportions of functional amino acids.

Conclusions and perspectives

In this review, we focused on research into the dose and dose intensity of the causes, models and nutritional strategies for oxidative stress in pigs. However, there are still many pressing issues that need to be addressed in the field of oxidative stress models and nutritional regulation

in pigs: (1) Ideal levels of ROS are essential for maintaining animal health, so we should first find the ranges of ROS levels in pigs that are most beneficial to their health before determining the appropriate amount of antioxidants in the diet; (2) At present, the main method used to determine whether pigs are under oxidative stress is to compare the relevant indexes of oxidative stress in the control group. In the future, existing data and new approaches should be used to determine the reference range of oxidative indexes; (3) A systemic comparison of the effects of different causes of oxidative stress should be carried out in the future; and (4) Nutritional strategies for oxidative stress in pigs should be thoroughly investigated. It is important to note that oxidative stress can reduce feed intake in pigs and it is unclear whether increasing nutrient density or changing the proportion of nutrients in the diet can eliminate the negative effects of oxidative stress. We hope that this review will serve as a theoretical basis and reference for the development of accurate models and nutritional strategies for specific oxidative stress.

Abbreviation

AFB1	Aflatoxin B1
BW	Body weight
CAT	Catalase
CTX	Cyclophosphamide
DON	Deoxynivalenol
EUFs	<i>Eucommia ulmoides</i> flavones
G	Gestation
GCLC	Glutamate cysteine ligase catalytic
GCLM	Glutamate cysteine ligase modifier
GPx	Glutathione peroxidase
GSH	Glutathione
GSR	Glutathione reductase
GSSG	Glutathione oxidized
GSTA	Glutathione S-transferase A
4-HNE	4-Hydroxynonenal
HO-1	Heme oxygenase 1
H ₂ O ₂	Hydrogen peroxide
IPEC	Intestinal porcine epithelial cell line
IUGR	Intrauterine growth restriction
L	Lactation
LPS	Lipopolysaccharide
MDA	Malondialdehyde
MLNs	Mesenteric lymph nodes
NAC	N-Acetylcysteine
NF-κB	Nuclear factor-κB
NO	Nitric oxide
NQO1	NAD(P)H:quinone oxidoreductase 1
Nrf2	Nuclear factor erythroid 2-related factor 2
8-OHG	8-Hydroxyguanosine
8-OHdG	8-Hydroxy-2 deoxyguanosine
OTA	Ochratoxin A
PMECs	Porcine mammary epithelial cells
RLU	Relative light unit
ROS	Reactive oxygen species
SOD	Superoxide dismutase
STAT	Signal transducer and activator of transcription
T-AOC	Total antioxidant capacity
TBARS	Thiobarbituric acid reactive substances
UCP2	Uncoupling protein 2
ZO-1	Zonula occludens-1

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Authors' contributions

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Declarations

Ethics approval and consent to participate

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The authors declare that they have no competing interests.

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References

- Sies H, Berndt C, Jones DP. Oxidative stress. *Annu Rev Biochem*. 2017;86:715–48.
- Yang X, Hu R, Shi M, Wang L, Yan J, Gong J, et al. Placental malfunction, fetal survival and development caused by sow metabolic disorder: the impact of maternal oxidative stress. *Antioxidants (Basel)*. 2023;12:360. <https://doi.org/10.3390/antiox12020360>.
- Campbell JM, Crenshaw JD, Polo J. The biological stress of early weaned piglets. *J Anim Sci Biotechnol*. 2013;4:19. <https://doi.org/10.1186/2049-1891-4-19>.
- Hao Y, Xing M, Gu X. Research progress on oxidative stress and its nutritional regulation strategies in pigs. *Animals (Basel)*. 2021;11:1384. <https://doi.org/10.3390/ani11051384>.
- Ao J, Zheng L, Peng J. Impact of progressive oxidative stress on sow reproductive performance and its nutritional impact of progressive oxidative stress on sow reproductive performance and its nutritional manipulation. *Chin J Animal Nutr*. 2016;28(12):3735–41. <https://doi.org/10.3969/j.issn.1006-267x.2016.12.003>.
- Berchieri-Ronchi CB, Kim SW, Zhao Y, Correa CR, Yeum K-J, Ferreira ALA. Oxidative stress status of highly prolific sows during gestation and lactation. *Animal*. 2011;5:1774–9. <https://doi.org/10.1017/S175173111000772>.
- Tan C, Wei H, Sun H, Ao J, Long G, Jiang S, et al. Effects of dietary supplementation of oregano essential oil to sows on oxidative stress status, lactation feed intake of sows, and piglet performance. *Biomed Res Int*. 2015;2015:525218. <https://doi.org/10.1155/2015/525218>.
- Jain L. Stress at birth and its inextricable link to the neonatal transition. *Obstet Gynecol*. 2016;128:685–7. <https://doi.org/10.1097/AOG.0000000000001657>.
- Yin J, Ren W, Liu G, Duan J, Yang G, Wu L, et al. Birth oxidative stress and the development of an antioxidant system in newborn piglets. *Free Radic Res*. 2013;47:1027–35. <https://doi.org/10.3109/10715762.2013.848277>.

10. Gao H, Chen X, Zhao J, Xue Z, Zhang L, Zhao F, et al. Integrative analysis of liver metabolomics and transcriptomics reveals oxidative stress in piglets with intrauterine growth restriction. *Biology (Basel)*. 2022. <https://doi.org/10.3390/biology11101430>.
11. Wjitten PJA, van der Meulen J, Verstegen MWA. Intestinal barrier function and absorption in pigs after weaning: a review. *Br J Nutr*. 2011;105:967–81. <https://doi.org/10.1017/S0007114510005660>.
12. Wei X, Tsai T, Howe S, Zhao J. Weaning induced gut dysfunction and nutritional interventions in nursery pigs: a partial review. *Animals (Basel)*. 2021;11:1279. <https://doi.org/10.3390/ani11051279>.
13. Zheng L, Duarte ME, Sevarolli Loftus A, Kim SW. Intestinal health of pigs upon weaning: challenges and nutritional intervention. *Front Vet Sci*. 2021;8:628258. <https://doi.org/10.3389/fvets.2021.628258>.
14. Xiong X, Tan B, Song M, Ji P, Kim K, Yin Y, et al. Nutritional intervention for the intestinal development and health of weaned piglets. *Front Vet Sci*. 2019;6:46. <https://doi.org/10.3389/fvets.2019.00046>.
15. Yin J, Wu MM, Xiao H, Ren WK, Duan JL, Yang G, et al. Development of an antioxidant system after early weaning in piglets. *J Anim Sci*. 2014;92:612–9. <https://doi.org/10.2527/jas.2013-6986>.
16. Cao ST, Wang CC, Wu H, Zhang QH, Jiao LF, Hu CH. Weaning disrupts intestinal antioxidant status, impairs intestinal barrier and mitochondrial function, and triggers mitophagy in piglets. *J Anim Sci*. 2018;96:1073–83. <https://doi.org/10.1093/jas/skx062>.
17. Liu F, Zhao W, Le HH, Cottrell JJ, Green MP, Leury BJ, et al. Review: What have we learned about the effects of heat stress on the pig industry? *Animal*. 2022;16(Suppl 2):100349. <https://doi.org/10.1016/j.animal.2021.100349>.
18. Cottrell JJ, Liu F, Hung AT, DiGiacomo K, Chauhan SS, Leury BJ, et al. Nutritional strategies to alleviate heat stress in pigs. *Anim Prod Sci*. 2015;55:1391. <https://doi.org/10.1071/AN15255>.
19. Zhang D, Wang L, Ma S, Ma H, Di L. Characterization of pig skeletal muscle transcriptomes in response to low temperature. *Vet Med Sci*. 2023;9:181–90. <https://doi.org/10.1002/vms3.1025>.
20. Liu L, Tai M, Yao W, Zhao R, Shen M. Effects of heat stress on posture transitions and reproductive performance of primiparous sows during late gestation. *J Therm Biol*. 2021;96:102828. <https://doi.org/10.1016/j.jtherbio.2020.102828>.
21. Zhao W, Liu F, Marth CD, Green MP, Le HH, Leury BJ, et al. Maternal heat stress alters expression of genes associated with nutrient transport activity and metabolism in female placentae from mid-gestating pigs. *Int J Mol Sci*. 2021. <https://doi.org/10.3390/ijms22084147>.
22. Zhao Y, Huang Y, Gao K, Wen X, Hu S, Wang L, et al. Maternal resveratrol regulates the growth performance, antioxidant capacity, and intestinal health of suckling piglets through intestinal microorganisms at high summer temperatures. *Front Nutr*. 2022;9:971496. <https://doi.org/10.3389/fnut.2022.971496>.
23. Guo H, He J, Yang X, Zheng W, Yao W. Responses of intestinal morphology and function in offspring to heat stress in primiparous sows during late gestation. *J Therm Biol*. 2020;89:102539. <https://doi.org/10.1016/j.jtherbio.2020.102539>.
24. Muns R, Malmkvist J, Larsen MLV, Sørensen D, Pedersen LJ. High environmental temperature around farrowing induced heat stress in crated sows. *J Anim Sci*. 2016;94:377–84. <https://doi.org/10.2527/jas.2015-9623>.
25. Zhao W, Artaiz O, Iqbal Y, Le HH, DiGiacomo K, Leury BJ, et al. Heat stress of gilts around farrowing causes oxygen insufficiency in the umbilical cord and reduces piglet survival. *Animal*. 2022;16:100668. <https://doi.org/10.1016/j.animal.2022.100668>.
26. Black JL, Mullan BP, Lorschly ML, Giles LR. Lactation in the sow during heat stress. *Livest Prod Sci*. 1993;35:153–70. [https://doi.org/10.1016/0301-6226\(93\)90188-N](https://doi.org/10.1016/0301-6226(93)90188-N).
27. Montilla SIR, Johnson TP, Pearce SC, Gardan-Salmon D, Gabler NK, Ross JW, et al. Heat stress causes oxidative stress but not inflammatory signaling in porcine skeletal muscle. *Temperature (Austin)*. 2014;1:42–50. <https://doi.org/10.4161/temp.28844>.
28. Brownstein AJ, Ganesan S, Summers CM, Pearce S, Hale BJ, Ross JW, et al. Heat stress causes dysfunctional autophagy in oxidative skeletal muscle. *Physiol Rep*. 2017;5:e13317. <https://doi.org/10.14814/phy2.13317>.
29. Ganesan S, Reynolds C, Hollinger K, Pearce SC, Gabler NK, Baumgard LH, et al. Twelve hours of heat stress induces inflammatory signaling in porcine skeletal muscle. *Am J Physiol Regul Integr Comp Physiol*. 2016;310:R1288–96. <https://doi.org/10.1152/ajpregu.00494.2015>.
30. Blecha F, Kelley KW. Cold stress reduces the acquisition of colostral immunoglobulin in piglets. *J Anim Sci*. 1981;52:594–600. <https://doi.org/10.2527/jas1981.523594x>.
31. Yu J, Chen S, Zeng Z, Xing S, Chen D, Yu B, et al. Effects of cold exposure on performance and skeletal muscle fiber in weaned piglets. *Animals (Basel)*. 2021. <https://doi.org/10.3390/ani11072148>.
32. Yang C, Cao C, Liu J, Zhao Y, Pan J, Tao C, et al. Distinct transcriptional responses of skeletal muscle to short-term cold exposure in Tibetan pigs and Bama pigs. *Int J Mol Sci*. 2023. <https://doi.org/10.3390/ijms24087431>.
33. Zhang Y, Sun L, Zhu R, Zhang S, Liu S, Wang Y, et al. Porcine gut microbiota in mediating host metabolic adaptation to cold stress. *NPJ Biofilms Microbiomes*. 2022;8:18. <https://doi.org/10.1038/s41522-022-00283-2>.
34. Teng T, Sun G, Ding H, Song X, Bai G, Shi B, et al. Characteristics of glucose and lipid metabolism and the interaction between gut microbiota and colonic mucosal immunity in pigs during cold exposure. *J Anim Sci Biotechnol*. 2023;14:84. <https://doi.org/10.1186/s40104-023-00886-5>.
35. Xiao H, Wu MM, Tan BE, Yin YL, Li TJ, Xiao DF, et al. Effects of composite antimicrobial peptides in weaning piglets challenged with deoxyvalenol: I. Growth performance, immune function, and antioxidation capacity. *J Anim Sci*. 2013;91:4772–80. <https://doi.org/10.2527/jas.2013-6426>.
36. Qiu Y, Nie X, Yang J, Wang L, Zhu C, Yang X, et al. Effect of resveratrol supplementation on intestinal oxidative stress, immunity and gut microbiota in weaned piglets challenged with deoxyvalenol. *Antioxidants (Basel)*. 2022. <https://doi.org/10.3390/antiox11091775>.
37. Qiu Y, Yang J, Wang L, Yang X, Gao K, Zhu C, et al. Dietary resveratrol attenuation of intestinal inflammation and oxidative damage is linked to the alteration of gut microbiota and butyrate in piglets challenged with deoxyvalenol. *J Anim Sci Biotechnol*. 2021;12:71. <https://doi.org/10.1186/s40104-021-00596-w>.
38. Marin DE, Pistol GC, Gras M, Palade M, Taranu I. A comparison between the effects of ochratoxin A and aristolochic acid on the inflammation and oxidative stress in the liver and kidney of weaning piglets. *Naunyn Schmiedeberg Arch Pharmacol*. 2018;391:1147–56. <https://doi.org/10.1007/s00210-018-1538-9>.
39. Marin DE, Bulgaru CV, Anghel CA, Pistol GC, Dore MI, Palade ML, et al. Grape seed waste counteracts aflatoxin B1 toxicity in piglet mesenteric lymph nodes. *Toxins (Basel)*. 2020;12:800. <https://doi.org/10.3390/toxins12120800>.
40. Wu L, Liao P, He L, Ren W, Yin J, Duan J, et al. Growth performance, serum biochemical profile, jejunal morphology, and the expression of nutrients transporter genes in deoxyvalenol (DON)-challenged growing pigs. *BMC Vet Res*. 2015;11:144. <https://doi.org/10.1186/s12917-015-0449-y>.
41. Wu L, Wang W, Yao K, Zhou T, Yin J, Li T, et al. Effects of dietary arginine and glutamine on alleviating the impairment induced by deoxyvalenol stress and immune relevant cytokines in growing pigs. *PLOS One*. 2013;8:e69502. <https://doi.org/10.1371/journal.pone.0069502>.
42. Rosero DS, Odle J, Moeser AJ, Boyd RD, van Heugten E. Peroxidised dietary lipids impair intestinal function and morphology of the small intestine villi of nursery pigs in a dose-dependent manner. *Br J Nutr*. 2015;114:1985–92. <https://doi.org/10.1017/S000711451500392X>.
43. Huang L, Ma X-Y, Jiang Z-Y, Hu Y-J, Zheng C-T, Yang X-F, et al. Effects of soybean isoflavone on intestinal antioxidant capacity and cytokines in young piglets fed oxidized fish oil. *J Zhejiang Univ Sci B*. 2016;17:965–74. <https://doi.org/10.1631/jzus.B1600078>.
44. Wu L, Li J, Li Y, Li T, He Q, Tang Y, et al. Aflatoxin B1, zearalenone and deoxyvalenol in feed ingredients and complete feed from different province in China. *J Anim Sci Biotechnol*. 2016;7:63. <https://doi.org/10.1186/s40104-016-0122-8>.
45. Tian M, Feng Y, He X, Zhang D, Wang W, Di L. Mycotoxins in live-stock feed in China - Current status and future challenges. *Toxicon*. 2022;214:112–20. <https://doi.org/10.1016/j.toxicon.2022.05.041>.
46. Adegbeye MJ, Reddy PRK, Chilaka CA, Balogun OB, Elghandour MMMY, Rivas-Caceres RR, et al. Mycotoxin toxicity and residue in animal products: prevalence, consumer exposure and reduction strategies - a review. *Toxicon*. 2020;177:96–108. <https://doi.org/10.1016/j.toxicon.2020.01.007>.

47. Magnoli AP, Poloni VL, Cavaglieri L. Impact of mycotoxin contamination in the animal feed industry. *Curr Opin Food Sci.* 2019;29:99–108. <https://doi.org/10.1016/j.cofs.2019.08.009>.
48. Xu R, Kiarie EG, Yiannikouris A, Sun L, Karrow NA. Nutritional impact of mycotoxins in food animal production and strategies for mitigation. *J Anim Sci Biotechnol.* 2022;13:69. <https://doi.org/10.1186/s40104-022-00714-2>.
49. Liu S, Li J, Kang W, Li Y, Ge L, Liu D, et al. Aflatoxin B1 induces intestinal barrier dysfunction by regulating the FXR-mediated MLCK signaling pathway in mice and in IPEC-J2 cells. *J Agric Food Chem.* 2023;71:867–76. <https://doi.org/10.1021/acs.jafc.2c06931>.
50. Seppanen CM, Saari CA. Formation of 4-hydroxynonenal, a toxic aldehyde, in soybean oil at frying temperature. *J Amer Oil Chem Soc.* 2002;79:1033–8. <https://doi.org/10.1007/s11746-002-0598-z>.
51. Ayala A, Muñoz MF, Argüelles S. Lipid peroxidation: Production, metabolism, and signaling mechanisms of malondialdehyde and 4-hydroxy-2-nonenal. *Oxid Med Cell Longev.* 2014;2014:360438. <https://doi.org/10.1155/2014/360438>.
52. Chang PL, Boyd RD, Zier-Rush C, Rosero DS, van Heugten E. Lipid peroxidation impairs growth and viability of nursery pigs reared under commercial conditions. *J Anim Sci.* 2019;97:3379–89. <https://doi.org/10.1093/jas/skz183>.
53. Ji Y, Dai Z, Wu G, Wu Z. 4-Hydroxy-2-nonenal induces apoptosis by activating ERK1/2 signaling and depleting intracellular glutathione in intestinal epithelial cells. *Sci Rep.* 2016;6:32929. <https://doi.org/10.1038/srep32929>.
54. Liang H, Liu N, Wang R, Zhang Y, Chen J, Dai Z, et al. N-acetyl serotonin alleviates oxidative damage by activating nuclear factor erythroid 2-related factor 2 signaling in porcine enterocytes. *Antioxidants (Basel).* 2020;9:303. <https://doi.org/10.3390/antiox9040303>.
55. Liu N, Ma X, Luo X, Zhang Y, He Y, Dai Z, et al. L-glutamine attenuates apoptosis in porcine enterocytes by regulating glutathione-related redox homeostasis. *J Nutr.* 2018;148:526–34. <https://doi.org/10.1093/jn/nxx062>.
56. Ji Y, He Y, Yang Y, Dai Z, Wu Z. Hydroxyproline alleviates 4-hydroxy-2-nonenal-induced DNA damage and apoptosis in porcine intestinal epithelial cells. *Anim Nutr.* 2022;9:7–15. <https://doi.org/10.1016/j.aninu.2021.08.003>.
57. Yin J, Liu M, Ren W, Duan J, Yang G, Zhao Y, et al. Effects of dietary supplementation with glutamate and aspartate on diquat-induced oxidative stress in piglets. *PLoS One.* 2015;10:e0122893. <https://doi.org/10.1371/journal.pone.0122893>.
58. Lv M, Yu B, Mao XB, Zheng P, He J, Chen DW. Responses of growth performance and tryptophan metabolism to oxidative stress induced by diquat in weaned pigs. *Animal.* 2012;6:928–34. <https://doi.org/10.1017/S1751731110002382>.
59. Cao S, Wu H, Wang C, Zhang Q, Jiao L, Lin F, et al. Diquat-induced oxidative stress increases intestinal permeability, impairs mitochondrial function, and triggers mitophagy in piglets. *J Anim Sci.* 2018;96:1795–805. <https://doi.org/10.1093/jas/sky104>.
60. Duan J, Yin J, Ren W, Liu T, Cui Z, Huang X, et al. Dietary supplementation with L-glutamate and L-aspartate alleviates oxidative stress in weaned piglets challenged with hydrogen peroxide. *Amino Acids.* 2016;48:53–64. <https://doi.org/10.1007/s00726-015-2065-3>.
61. Yi D, Hou Y, Wang L, Ding B, Yang Z, Li J, et al. Dietary N-acetylcysteine supplementation alleviates liver injury in lipopolysaccharide-challenged piglets. *Br J Nutr.* 2014;111:46–54. <https://doi.org/10.1017/S0007114513002171>.
62. Cao S, Zhang Q, Wang C, Wu H, Jiao L, Hong Q, et al. LPS challenge increased intestinal permeability, disrupted mitochondrial function and triggered mitophagy of piglets. *Innate Immun.* 2018;24:221–30. <https://doi.org/10.1177/1753425918769372>.
63. Zhou Y, Hu X, Zhong S, Yu W, Wang J, Zhu W, et al. Effects of continuous LPS induction on oxidative stress and liver injury in weaned piglets. *Vet Sci.* 2022;10:22. <https://doi.org/10.3390/vetsci10010022>.
64. Lee SI, Kang KS. Omega-3 fatty acids modulate cyclophosphamide induced markers of immunosuppression and oxidative stress in pigs. *Sci Rep.* 2019;9:2684. <https://doi.org/10.1038/s41598-019-39458-x>.
65. Xu J, Yu B, Chen D. Duration and indicators of oxidation stress induced by diquat in growing pigs. *Scientia Agricultura Sinica.* 2008;41(12):4359–64. <https://doi.org/10.3864/j.issn.0578-1752.2008.12.056>.
66. Xu YQ, Xing YY, Wang ZQ, Yan SM, Shi BL. Pre-protective effects of dietary chitosan supplementation against oxidative stress induced by diquat in weaned piglets. *Cell Stress Chaperones.* 2018;23:703–10. <https://doi.org/10.1007/s12192-018-0882-5>.
67. Zou Y, Wang J, Peng J, Wei H. Oregon essential oil induces SOD1 and GSH expression through Nrf2 activation and alleviates hydrogen peroxide-induced oxidative damage in IPEC-J2 cells. *Oxid Med Cell Longev.* 2016;2016:5987183. <https://doi.org/10.1155/2016/5987183>.
68. Xiao H, Wu M, Shao F, Guan G, Huang B, Tan B, et al. N-Acetyl-L-cysteine protects the enterocyte against oxidative damage by modulation of mitochondrial function. *Mediators Inflamm.* 2016;2016:8364279. <https://doi.org/10.1155/2016/8364279>.
69. Yin J, Duan J, Cui Z, Ren W, Li T, Yin Y. Hydrogen peroxide-induced oxidative stress activates NF- κ B and Nrf2/Keap1 signals and triggers autophagy in piglets. *RSC Adv.* 2015;5:15479–86. <https://doi.org/10.1039/C4RA13557A>.
70. Yin J, Wu M, Li Y, Ren W, Xiao H, Chen S, et al. Toxicity assessment of hydrogen peroxide on Toll-like receptor system, apoptosis, and mitochondrial respiration in piglets and IPEC-J2 cells. *Oncotarget.* 2017;8:3124–31. <https://doi.org/10.18632/oncotarget.13844>.
71. Cao S, Xiao H, Li X, Zhu J, Gao J, Wang L, et al. AMPK-PINK1/Parkin mediated mitophagy is necessary for alleviating oxidative stress-induced intestinal epithelial barrier damage and mitochondrial energy metabolism dysfunction in IPEC-J2. *Antioxidants (Basel).* 2021;10:2010. <https://doi.org/10.3390/antiox10122010>.
72. Batista CRA, Gomes GF, Candelario-Jalil E, Fiebich BL, de Oliveira ACP. Lipopolysaccharide-induced neuroinflammation as a bridge to understand neurodegeneration. *Int J Mol Sci.* 2019;20:2293. <https://doi.org/10.3390/ijms20092293>.
73. Lugin J, Rosenblatt-Velin N, Parapanov R, Liaudet L. The role of oxidative stress during inflammatory processes. *Biol Chem.* 2014;395:203–30. <https://doi.org/10.1515/hsz-2013-0241>.
74. Chen F, Wang H, Chen J, Liu Y, Wen W, Li Y, et al. *Lactobacillus delbrueckii* ameliorates intestinal integrity and antioxidant ability in weaned piglets after a lipopolysaccharide challenge. *Oxid Med Cell Longev.* 2020;2020:6028606. <https://doi.org/10.1155/2020/6028606>.
75. Lee SI, Kang KS. Function of capric acid in cyclophosphamide-induced intestinal inflammation, oxidative stress, and barrier function in pigs. *Sci Rep.* 2017;7:16530. <https://doi.org/10.1038/s41598-017-16561-5>.
76. Han H, Liu Z, Yin J, Gao J, He L, Wang C, et al. D-galactose induces chronic oxidative stress and alters gut microbiota in weaned piglets. *Front Physiol.* 2021;12:634283. <https://doi.org/10.3389/fphys.2021.634283>.
77. Chen J, Huang Z, Cao X, Chen X, Zou T, You J. Plant-derived polyphenols as Nrf2 activators to counteract oxidative stress and intestinal toxicity induced by deoxynivalenol in swine: an emerging research direction. *Antioxidants (Basel).* 2022;11:2379. <https://doi.org/10.3390/antiox11122379>.
78. Garcia-Contreras C, Vazquez-Gomez M, Barbero A, Pesantez JL, Zinellu A, Berlinguer F, et al. Polyphenols and IUGR pregnancies: effects of maternal hydroxytyrosol supplementation on placental gene expression and fetal antioxidant status, DNA-methylation and phenotype. *Int J Mol Sci.* 2019;20:1187. <https://doi.org/10.3390/ijms20051187>.
79. Yin J, Han H, Liu Z, He X, Li T, Yin Y. Regulation of functional amino acids on oxidative stress in pigs. *Sci Sin-Vitae.* 2019;49:193–201. <https://doi.org/10.1360/N052018-00159>.
80. Scott MB, Styring AK, McCullagh JSO. Polyphenols: bioavailability, microbiome interactions and cellular effects on health in humans and animals. *Pathogens.* 2022;11:770. <https://doi.org/10.3390/pathogens11070770>.
81. Gessner DK, Ringseis R, Eder K. Potential of plant polyphenols to combat oxidative stress and inflammatory processes in farm animals. *J Anim Physiol Anim Nutr (Berl).* 2017;101:605–28. <https://doi.org/10.1111/jpn.12579>.
82. Li Y, Jiang X, Cai L, Zhang Y, Ding H, Yin J, et al. Effects of daidzein on antioxidant capacity in weaned pigs and IPEC-J2 cells. *Anim Nutr.* 2022;11:48–59. <https://doi.org/10.1016/j.aninu.2022.06.014>.
83. Chen W, Ma X, Lin Y, Xiong Y, Zheng C, Hu Y, et al. Dietary supplementation with a high dose of daidzein enhances the antioxidant capacity in swine muscle but experts pro-oxidant function in liver and fat tissues. *J Anim Sci Biotechnol.* 2016;7:43. <https://doi.org/10.1186/s40104-016-0102-z>.

84. Li S, Wen X, Jiang Z, Wang L, Gao K, Xiao H. Effects of dietary supplementation of isoflavone-S in late gestation and lactation on reproductive performance, antioxidant capacity and immune functions of sows. *Chin J Anim Nutr.* 2022;34(10):6459–68. <https://doi.org/10.3969/j.issn.1006-267x.2022.10.041>.
85. Li Y, He G, Chen D, Yu B, Yu J, Zheng P, et al. Supplementing daidzein in diets improves the reproductive performance, endocrine hormones and antioxidant capacity of multiparous sows. *Anim Nutr.* 2021;7:1052–60. <https://doi.org/10.1016/j.aninu.2021.09.002>.
86. Xun W, Fu Q, Shi L, Cao T, Jiang H, Ma Z. Resveratrol protects intestinal integrity, alleviates intestinal inflammation and oxidative stress by modulating AhR/Nrf2 pathways in weaned piglets challenged with diquat. *Int Immunopharmacol.* 2021;99:107989. <https://doi.org/10.1016/j.intimp.2021.107989>.
87. Cao S, Shen Z, Wang C, Zhang Q, Hong Q, He Y, et al. Resveratrol improves intestinal barrier function, alleviates mitochondrial dysfunction and induces mitophagy in diquat challenged piglets. *Food Funct.* 2019;10:344–54. <https://doi.org/10.1039/c8fo02091d>.
88. Gao Y, Meng Q, Qin J, Zhao Q, Shi B. Resveratrol alleviates oxidative stress induced by oxidized soybean oil and improves gut function via changing gut microbiota in weaned piglets. *J Anim Sci Biotechnol.* 2023;14:54. <https://doi.org/10.1186/s40104-023-00851-2>.
89. Meng Q, Guo T, Li G, Sun S, He S, Cheng B, et al. Dietary resveratrol improves antioxidant status of sows and piglets and regulates antioxidant gene expression in placenta by Keap1-Nrf2 pathway and Sirt1. *J Anim Sci Biotechnol.* 2018;9:34. <https://doi.org/10.1186/s40104-018-0248-y>.
90. Zhang H, Chen Y, Chen Y, Jia P, Ji S, Xu J, et al. Comparison of the effects of resveratrol and its derivative pterostilbene on hepatic oxidative stress and mitochondrial dysfunction in piglets challenged with diquat. *Food Funct.* 2020;11:4202–15. <https://doi.org/10.1039/d0fo00732c>.
91. Chen Y, Zhang H, Chen Y, Jia P, Ji S, Zhang Y, et al. Resveratrol and its derivative pterostilbene ameliorate intestine injury in intrauterine growth-retarded weanling piglets by modulating redox status and gut microbiota. *J Anim Sci Biotechnol.* 2021;12:70. <https://doi.org/10.1186/s40104-021-00589-9>.
92. Niu Y, He J, Ahmad H, Shen M, Zhao Y, Gan Z, et al. Dietary curcumin supplementation increases antioxidant capacity, upregulates Nrf2 and Hmox1 levels in the liver of piglet model with intrauterine growth retardation. *Nutrients.* 2019;11:2978. <https://doi.org/10.3390/nu11122978>.
93. Zhang L, Zhang J, Yan E, He J, Zhong X, Zhang L, et al. Dietary supplemented curcumin improves meat quality and antioxidant status of intrauterine growth retardation growing pigs via Nrf2 signal pathway. *Animals (Basel).* 2020;10:539. <https://doi.org/10.3390/ani10030539>.
94. Yan E, Zhang J, Han H, Wu J, Gan Z, Wei C, et al. Curcumin alleviates IUGR jejunum damage by increasing antioxidant capacity through Nrf2/Keap1 pathway in growing pigs. *Animals (Basel).* 2019;10:41. <https://doi.org/10.3390/ani10010041>.
95. Zhang J, Yan E, Zhang L, Wang T, Wang C. Curcumin reduces oxidative stress and fat deposition in longissimus dorsi muscle of intrauterine growth-retarded finishing pigs. *Anim Sci J.* 2022;93:e13741. <https://doi.org/10.1111/asj.13741>.
96. Yuan D, Hussain T, Tan B, Liu Y, Ji P, Yin Y. The evaluation of antioxidant and anti-inflammatory effects of Eucommia ulmoides flavones using diquat-challenged piglet models. *Oxid Med Cell Longev.* 2017;2017:8140962. <https://doi.org/10.1155/2017/8140962>.
97. Xiao D, Yuan D, Tan B, Wang J, Liu Y, Tan B. The role of Nrf2 signaling pathway in Eucommia ulmoides flavones regulating oxidative stress in the intestine of piglets. *Oxid Med Cell Longev.* 2019;2019:9719618. <https://doi.org/10.1155/2019/9719618>.
98. Hong C, Gao K, Wang L, Xiao H. Application of isoflavones and their metabolic derivatives in application of isoflavones and their metabolic derivatives in livestock and poultry production. *Chin J Anim Nutr.* 2023;35(6):3445–55. <https://doi.org/10.12418/CJAN2023.320>.
99. Meng Q, Li J, Wang C, Shan A. Biological function of resveratrol and its application in animal production: a review. *J Anim Sci Biotechnol.* 2023;14:25. <https://doi.org/10.1186/s40104-022-00822-z>.
100. Yang J, Zhu C, Ye J, Lv Y, Wang L, Chen Z, et al. Protection of porcine intestinal-epithelial cells from deoxynivalenol-induced damage by resveratrol via the Nrf2 signaling pathway. *J Agric Food Chem.* 2019;67:1726–35. <https://doi.org/10.1021/acs.jafc.8b03662>.
101. Huang Y, Zheng C, Song B, Wang L, Xiao H, Jiang Z. Resveratrol ameliorates intestinal damage challenged with deoxynivalenol through mitophagy in vitro and in vivo. *Front Vet Sci.* 2021;8:807301. <https://doi.org/10.3389/fvets.2021.807301>.
102. Li Y, Zhang H, Tu F, Cao J, Hou X, Chen Y, et al. Effects of resveratrol and its derivative pterostilbene on hepatic injury and immunological stress of weaned piglets challenged with lipopolysaccharide. *J Anim Sci.* 2022. <https://doi.org/10.1093/jas/skac339>.
103. Ahmed ST, Hossain ME, Kim GM, Hwang JA, Ji H, Yang CJ. Effects of resveratrol and essential oils on growth performance, immunity, digestibility and fecal microbial shedding in challenged piglets. *Asian Australas J Anim Sci.* 2013;26:683–90. <https://doi.org/10.5713/ajas.2012.12683>.
104. Tang X, Xiong K, Wessie T, Wu X. Curcumin and intestinal oxidative stress of pigs with intrauterine growth retardation: a review. *Front Nutr.* 2022;9:847673. <https://doi.org/10.3389/fnut.2022.847673>.
105. Cao S, Wang C, Yan J, Li X, Wen J, Hu C. Curcumin ameliorates oxidative stress-induced intestinal barrier injury and mitochondrial damage by promoting Parkin dependent mitophagy through AMPK-TFEB signal pathway. *Free Radic Biol Med.* 2020;147:8–22. <https://doi.org/10.1016/j.freeradbiomed.2019.12.004>.
106. Li X, Zhu J, Lin Q, Yu M, Lu J, Feng J, et al. Effects of curcumin on mitochondrial function, endoplasmic reticulum stress, and mitochondria-associated endoplasmic reticulum membranes in the jejunum of oxidative stress piglets. *J Agric Food Chem.* 2022;70:8974–85. <https://doi.org/10.1021/acs.jafc.2c02824>.
107. Hussain T, Tan B, Liu G, Oladele OA, Rahu N, Tossou MC, et al. Health-promoting properties of Eucommia ulmoides: a review. *Evid Based Complement Alternat Med.* 2016;2016:5202908. <https://doi.org/10.1155/2016/5202908>.
108. Wu G. Functional amino acids in growth, reproduction, and health. *Adv Nutr.* 2010;1:31–7. <https://doi.org/10.3945/an.110.1008>.
109. Wu G. Functional amino acids in nutrition and health. *Amino Acids.* 2013;45:407–11. <https://doi.org/10.1007/s00726-013-1500-6>.
110. Ma X, Lin Y, Jiang Z, Zheng C, Zhou G, Yu D, et al. Dietary arginine supplementation enhances antioxidative capacity and improves meat quality of finishing pigs. *Amino Acids.* 2010;38:95–102. <https://doi.org/10.1007/s00726-008-0213-8>.
111. Zheng P, Yu B, He J, Tian G, Luo Y, Mao X, et al. Protective effects of dietary arginine supplementation against oxidative stress in weaned piglets. *Br J Nutr.* 2013;109:2253–60. <https://doi.org/10.1017/S0007114512004321>.
112. Wu G, Bazer FW, Satterfield MC, Li X, Wang X, Johnson GA, et al. Impacts of arginine nutrition on embryonic and fetal development in mammals. *Amino Acids.* 2013;45:241–56. <https://doi.org/10.1007/s00726-013-1515-z>.
113. Li J, Xia H, Yao W, Wang T, Piao X, Thacker P, et al. Effects of arginine supplementation during early gestation (day 1 to 30) on litter size and plasma metabolites in gilts and sows. *J Anim Sci.* 2015;93:5291–303. <https://doi.org/10.2527/jas.2014-8657>.
114. Correa-Aragunde N, Foresi N, Lamattina L. Nitric oxide is a ubiquitous signal for maintaining redox balance in plant cells: Regulation of ascorbate peroxidase as a case study. *J Exp Bot.* 2015;66:2913–21. <https://doi.org/10.1093/jxb/erv073>.
115. Qiu Y, Yang X, Wang L, Gao K, Jiang Z. L-arginine inhibited inflammatory response and oxidative stress induced by lipopolysaccharide via arginase-1 signaling in IPEC-J2 cells. *Int J Mol Sci.* 2019;20:1800. <https://doi.org/10.3390/ijms20071800>.
116. Li B, Li L, Donaldson PJ, Lim JC. Dynamic regulation of GSH synthesis and uptake pathways in the rat lens epithelium. *Exp Eye Res.* 2010;90:300–7. <https://doi.org/10.1016/j.exer.2009.11.006>.
117. Song Zh, Tong G, Xiao K, Le Jiao F, Ke Yi, Hu CH. L-cysteine protects intestinal integrity, attenuates intestinal inflammation and oxidant stress, and modulates NF- κ B and Nrf2 pathways in weaned piglets after LPS challenge. *Innate Immun.* 2016;22:152–61. <https://doi.org/10.1177/1753425916632303>.
118. Zhang H, Su W, Ying Z, Chen Y, Le Z, Li Y, et al. N-acetylcysteine attenuates intrauterine growth retardation-induced hepatic damage in suckling piglets by improving glutathione synthesis and cellular homeostasis. *Eur J Nutr.* 2018;57:327–38. <https://doi.org/10.1007/s00394-016-1322-x>.

119. Ni H, Long L, Bin P, Azad MAK, Xu K, Zhou X, et al. Maternal cysteine intake influenced oxidative status and lipid-related gut microbiota and plasma metabolomics in male suckling piglets. *Anim Feed Sci Technol*. 2021;276:114947. <https://doi.org/10.1016/j.anifeeds.2021.114947>.
120. Huang S, Zhang L, Luo J, Wu D, Ma K, Chen Y, et al. Cysteamine and N-Acetyl-cysteine alleviate placental oxidative stress and barrier function damage induced by deoxynivalenol. *J Agric Food Chem*. 2023;71:6846–58. <https://doi.org/10.1021/acs.jafc.3c00399>.
121. Hou Y, Wu G. L-Glutamate nutrition and metabolism in swine. *Amino Acids*. 2018;50:1497–510. <https://doi.org/10.1007/s00726-018-2634-3>.
122. Wu M, Xiao H, Ren W, Yin J, Hu J, Duan J, et al. An NMR-based metabolomic approach to investigate the effects of supplementation with glutamic acid in piglets challenged with deoxynivalenol. *PLoS One*. 2014;9:e113687. <https://doi.org/10.1371/journal.pone.0113687>.
123. Lv D, Xiong X, Yang H, Wang M, He Y, Liu Y, et al. Effect of dietary soy oil, glucose, and glutamine on growth performance, amino acid profile, blood profile, immunity, and antioxidant capacity in weaned piglets. *Sci China Life Sci*. 2018;61:1233–42. <https://doi.org/10.1007/s11427-018-9301-y>.
124. Wang W, Dai Z, Wu Z, Lin G, Jia S, Hu S, et al. Glycine is a nutritionally essential amino acid for maximal growth of milk-fed young pigs. *Amino Acids*. 2014;46:2037–45. <https://doi.org/10.1007/s00726-014-1758-3>.
125. Xu X, Wei Y, Hua H, Zhu H, Xiao K, Zhao J, et al. Glycine alleviated intestinal injury by inhibiting ferroptosis in piglets challenged with diquat. *Animals (Basel)*. 2022. <https://doi.org/10.3390/ani12223071>.
126. Wang W, Wu Z, Lin G, Hu S, Wang B, Dai Z, et al. Glycine stimulates protein synthesis and inhibits oxidative stress in pig small intestinal epithelial cells. *J Nutr*. 2014;144:1540–8. <https://doi.org/10.3945/jn.114.194001>.
127. Ji X, Tang Z, Zhang F, Zhou F, Wu Y, Wu D. Dietary taurine supplementation counteracts deoxynivalenol-induced liver injury via alleviating oxidative stress, mitochondrial dysfunction, apoptosis, and inflammation in piglets. *Ecotoxicol Environ Saf*. 2023;253:114705. <https://doi.org/10.1016/j.ecoenv.2023.114705>.
128. Xu M, Che L, Gao K, Wang L, Yang X, Wen X, et al. Effects of dietary taurine supplementation to gilts during late gestation and lactation on offspring growth and oxidative stress. *Animals (Basel)*. 2019. <https://doi.org/10.3390/ani9050220>.
129. Xu M, Che L, Gao K, Wang L, Yang X, Wen X, et al. Taurine alleviates oxidative stress in porcine mammary epithelial cells by stimulating the Nrf2-MAPK signaling pathway. *Food Sci Nutr*. 2023;11:1736–46. <https://doi.org/10.1002/fsn3.3203>.